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APPLICATION FOR LETTERS PATENT

for

**OLIGOPEPTIDE TREATMENT OF ANTHRAX**

Inventors:  
Nisar Asmed Khan  
Robert Benner

Attorney:  
Allen C. Turner  
Registration No. 33,041  
TRASKBRITT  
P.O. Box 2550  
Salt Lake City, Utah 84110  
(801) 532-1922

Title: OLIGOPEPTIDE TREATMENT OF ANTHRAX

#### CROSS-REFERENCE TO RELATED APPLICATION

This application is a continuation in part of U.S.S.N. 09/821,380 filed on March 29,  
5 2001, the contents of which are incorporated by this reference.

#### TECHNICAL FIELD

The invention relates to a method of treating disease, particularly anthrax.

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#### BACKGROUND

Anthrax, the disease caused by the spore-forming *Bacillus anthracis* (*B. anthracis*), continues to be a worldwide problem, among domesticated and wild herbivores in Asia and Africa and poses a worldwide threat when being used as biological weapons for biological warfare or bioterrorism. Human infections occur after contact with infected animals or  
15 contaminated animal products. Outbreaks or epidemics are a constant threat for endemic regions because spores can persist in the soil for long periods of time. Importation controls on certain animal products are necessary to prevent the establishment of anthrax where the disease is not endemic. Human anthrax is usually classified by the portal of entry into the host. Cutaneous anthrax, which accounts for the vast majority of human anthrax cases, is a  
20 localized infection with generally mild systemic symptoms and characterized by a painless papule that is surrounded by edema which can be quite extensive. The papule ulcerates by day 5 or 6 and develops into the characteristic black eschar of cutaneous anthrax. Inhalation anthrax, which occurs after inhaling airborne spores, gastrointestinal anthrax, resulting from ingestion of contaminated food, and in some instances, untreated cutaneous  
25 anthrax, are characterized by dissemination of the bacteria from the initial site of infection with development of a massive septicemia and toxemia. In inhalation anthrax, phagocytic cells transport the spores from the lung alveoli to the regional lymph nodes, where the spores germinate and bacteria multiply. The bacilli then spread into the bloodstream, where they are temporarily removed by the reticuloendothelial system. Prior to death, which  
30 occurs 2 to 5 days after infection, there is a sudden onset of acute symptoms characterized by hypotension, edema, and fatal shock due to an extensive septicemia and toxemia.

Therapeutic intervention in general must be initiated early, as septicemic infections are nearly always fatal.

The invention relates to the modulation of gene expression in a cell, also called gene control, in relation to the treatment of a variety of diseases such as anthrax. As said, anthrax is a disease of animals and humans, poses a significant threat as an agent of biological warfare and terrorism. Inhalational anthrax, in which spores of *B. anthracis* are inhaled, is almost always fatal, as diagnosis is rarely possible before the disease has progressed to a point where antibiotic treatment is ineffective. The major virulence factors of *B. anthracis* are a poly-D-glutamic acid capsule and anthrax toxin. Anthrax toxin consists of three distinct proteins that act in concert: two enzymes, lethal factor (LF) and edema factor (EF; an adenylate cyclase); and protective antigen (PA). The PA is a four-domain protein that binds a host cell-surface receptor by its carboxy-terminal domain; cleavage of its N-terminal domain by a furin-like protease allows PA to form heptamers that bind the toxic enzymes with high affinity through homologous N-terminal domains. The complex is endocytosed; acidification of the endosome leads to membrane insertion of the PA heptamer by forming a 14-stranded beta-barrel, followed by translocation of the toxic enzymes into the cytosol by an unknown mechanism. The binary combination of PA and LF is sufficient to induce rapid death in animals when given intravenously, and certain metalloprotease inhibitors block the effects of the toxin in vitro. Thus, LF is a potential target for therapeutic agents that would inhibit its catalytic activity or block its association with PA. LF is a protein (relative molecular mass 90,000) that is critical in the pathogenesis of anthrax. It comprises four domains: domain I binds the membrane-translocating component of anthrax toxin, the PA; domains II, III and IV together create a long deep groove that holds the 16-residue N-terminal tail of mitogen-activated protein kinase kinase-2 (MAPKK-2) before cleavage. Domain II resembles the ADP-ribosylating toxin from *Bacillus cereus*, but the active site has been mutated and recruited to augment substrate recognition. Domain III is inserted into domain II, and seems to have arisen from a repeated duplication of a structural element of domain II. Domain IV is distantly related to the zinc metalloprotease family, and contains the catalytic center; it also resembles domain I. The structure thus reveals a protein that has evolved through a process of gene duplication, mutation and fusion, into an enzyme with high and unusual specificity.

The MAPKK family of proteins are the only known cellular substrates of LF. Cleavage by LF near to their N termini removes the docking sequence for the downstream cognate MAP kinase. The effect of lethal toxin on tumor cells, for example, is to inhibit tumor growth and angiogenesis, most probably by inhibiting the MAPKK-1 and MAPKK-2 pathways. However, the primary cell type affected in anthrax pathogenesis is the macrophage. LF has been shown to cleave short N-terminal fragments from mitogen or extracellular signal-regulated MAPKK-1, MAPKK-2, MAPKK-3, and MAPKK-6, the upstream activators of extracellular signal-regulated kinase 1 (ERK1), ERK2, and p38. Recent data show that this results in inhibiting release, but not production, of the pro-inflammatory mediators, NO and tumor necrosis factor-alpha (TNF-alpha). In addition, high levels of lethal toxin lead to lysis of macrophages within a few hours, by an unknown mechanism. Recent data suggests that this happens due to inhibition of growth-factor pathways leading to macrophage death. These observations suggest that at an early stage in infection, lethal toxin may reduce (or delay) the immune response, whereas at a late stage in infection, high titers of the bacterium in the bloodstream trigger macrophage lysis and the sudden release of high levels of NO and TNF-alpha. This may explain the symptoms before death which are characterized by the hyperstimulation of host macrophage inflammatory pathways, leading to dramatic hypotension and shock. These symptoms resemble those of LPS induced septic shock, whereby it is of note LPS-nonresponder mice such as C3H/HeJ are also quite resistant against anthrax toxin.

The recognition sites for LF require the presence of the proline (P) residue followed by a hydrophobic residue or a glycine (G) residue, between which LF cleaves. The recognition sites further require an uncharged amino acid following the hydrophobic residue, and at least one positively charged amino acid (and no negatively charged amino acid, such as Asp and Glu) within the 5 amino acids to the N-terminal side of the proline residue. Other residues in the sequence provide appropriate spacing between the critical residues or between the donor and acceptor, and thus their composition is not critical, and can include any natural or unnatural amino acid.

Gene control is generally thought to occur at four levels: 1) transcription (either initiation or termination), 2) processing of primary transcripts, 3) stabilization or destabilization of mRNAs, and 4) mRNA translation. The primary function of gene control

in cells is to adjust the enzymatic machinery of the cell to its nutritional, chemical and physical environment.

It is generally thought that gene expression is regulated at both the level of transcription and translation. Modulation or regulation of gene expression requires factors called transcriptional factors. The term gene control or regulation also refers to the formation and use of mRNA. Although control can be exerted at a number of different molecular steps, differential transcription probably most frequently underlies the differential rate of protein synthesis in prokaryotes as well as eukaryotes. It is generally thought that activator proteins (also called transcription factors or transcriptional activators) bind to DNA and recruit the transcriptional machinery in a cell to a promoter, thereby stimulating gene expression. Further, differential processing of RNA transcripts in the cell nucleus, differential stabilization of mRNA in the cytoplasm, and differential translation of mRNA into protein are also important in eukaryotic gene control. These steps define the regulatory decisions in a transcriptional circuit and misregulation at any stage can result in a variety of diseases.

Where in unicellular organisms, be it of prokaryotic or eukaryotic origin, a cell's response to its environment is influenced by many stimuli from the outside world, reflecting the often widely variable environment of the single cell, most cells in multicellular organisms experience a fairly constant environment. Perhaps for this reason, genes that are devoted to responses to environmental changes constitute a much smaller fraction of the total number of genes in multicellular organisms than in single-cell organisms.

As said above, cells react to environmental changes, which they perceive through extracellular signals. These signals can be either physical (*e.g.*, light, temperature, pressure and electricity) or chemical (*e.g.*, food, hormones and neurotransmitters). Cells can both sense and produce signals. This makes it possible that they communicate with each other. In order to achieve this, there are complex signal-sensing and -producing mechanisms in uni- and multi-cellular organisms.

Two groups of chemical signals can be distinguished: membrane permeable and membrane-impermeable signals. The membrane-permeable signal molecules comprise the large family of steroid hormones, such as estrogens, progesterone and androgens. Steroids pass the plasma membrane and bind to specific receptors, which are localized in the

cytoplasm or nucleus of the cell. After binding of the hormone, the receptor undergoes a conformational change. The receptor is then able to bind to DNA itself or to proteins which can in turn interact with DNA. In general, steroid hormones can directly regulate gene expression by means of this process. The membrane-impermeable signal molecules include acetylcholine, growth factors, extracellular matrix components, thrombin, lysophosphatidic acid, the yeast mating factors and, for the social amoeba *Dictyostellium discoideum*, folic acid and cyclic AMP. They are recognized by receptors, which are localized on the plasma membrane of the cell. The receptors are specific for one particular signal molecule or a family of closely related signal molecules. Upon binding of their ligands, these receptors transduce the signals by several mechanisms.

The most characteristic and exacting requirement of gene control on multicellular organisms is the execution of precise developmental decisions so that the right gene is activated in the right cell at the right time. These developmental decisions include not only those related to the development of an organism per se, as for example can be seen during embryogenesis and organogenesis, or in response to disease but also relate to the differentiation or proliferation or apoptosis of those cells that merely carry out their genetic program essentially without leaving progeny behind.

Such cells, such as skin cells, precursors of red blood cells, lens cells of the eye, antibody-producing cells, are also often regulated by patterns of gene control that serve the need of the whole organism and not the survival of an individual cell.

It is generally reasoned that there are at least three components of gene control: molecular signals, levels and mechanisms. First, it is reasoned that specific signaling molecules exist to which a specific gene can respond. Second, control is exerted on one or more levels (*i.e.*, the step or steps) in the chain of events leading from the transcription of DNA to the use of mRNA in protein synthesis. Third, at each of those levels, specific molecular mechanisms are employed to finally exert the control over the gene to be expressed.

Many genes are regulated not by a signaling molecule that enters the cells but by molecules that bind to specific receptors on the surface of cells. Interaction between cell-surface receptors and their ligands can be followed by a cascade of intracellular events including variations in the intracellular levels of so-called second messengers (diacylglycerol,  $\text{Ca}^{2+}$ , cyclic nucleotides). The second messengers in turn lead to changes

in protein phosphorylation through the action of cyclic AMP, cyclic GMP, calcium-activated protein kinases, or protein kinase C, which is activated by diacylglycerol.

Many of the responses to binding of ligands to cell-surface receptors are cytoplasmatic and do not involve immediate gene activation in the nucleus. Some receptor-ligand interactions, however, are known to cause prompt nuclear transcriptional activation of a specific and limited set of genes. For example, one proto-oncogene, *c-fos*, is known to be activated in some cell types by elevation of almost every one of the known second messengers and also by at least two growth factors, platelet-derived growth factor and epidermal growth factor. However, progress has been slow in determining exactly how such activation is achieved. In a few cases, the transcriptional proteins that respond to cell-surface signals have been characterized.

One of the clearest examples of activation of a pre-existing inactive transcription factor following a cell-surface interaction is the nuclear factor (NF)-kappaB, which was originally detected because it stimulates the transcription of genes encoding immunoglobulins of the kappa class in B-lymphocytes. The binding site for NF-kappaB in the kappa gene is well defined (see, for example, P.A. Baeuerle and D.Baltimore, 1988, Science 242:540), providing an assay for the presence of the active factor. This factor exists in the cytoplasm of lymphocytes complexed with an inhibitor. Treatment of the isolated complex in vitro with mild denaturing conditions dissociates the complex, thus freeing NF-kappaB to bind to its DNA site. Release of active NF-kappaB in cells is now known to occur after a variety of stimuli including treating cells with bacterial lipopolysaccharide (LPS) and extracellular polypeptides as well as chemical molecules (e.g., phorbol esters) that stimulate intracellular phosphokinases. Thus a phosphorylation event triggered by many possible stimuli may account for NF-kappaB conversion to the active state. The active factor is then translocated to the cell nucleus to stimulate transcription only of genes with a binding site for active NF-kappaB.

The inflammatory response involves the sequential release of mediators and the recruitment of circulating leukocytes, which become activated at the inflammatory site and release further mediators (Nat. Med. 7:1294;2001). This response is self-limiting and resolves through the release of endogenous anti-inflammatory mediators and the clearance of inflammatory cells. The persistent accumulation and activation of leukocytes is a hallmark of chronic inflammation. Current approaches to the treatment of inflammation

rely on the inhibition of pro-inflammatory mediator production and of mechanisms that initiate the inflammatory response. However, the mechanisms by which the inflammatory response resolves might provide new targets in the treatment of chronic inflammation. Studies in different experimental models of resolving inflammation have identified several putative mechanisms and mediators of inflammatory resolution. We have shown that cyclopentenone prostaglandins (cyPGs) may be endogenous anti-inflammatory mediators and promote the resolution of inflammation *in vivo*. Others have shown a temporal shift to the production of anti-inflammatory lipoxins during the resolution of inflammation. In recent years, apoptosis has been identified as an important mechanism for the resolution of inflammation *in vivo*. It has been postulated that defects in leukocyte apoptosis are important in the pathogenesis of inflammatory disease. In addition, the selective induction of apoptosis in leukocytes may offer a new therapeutic approach to inflammatory disease.

Considering that NF-kappaB is thought by many to be a primary effector of disease (A.S. Baldwin, J. Clin. Invest., 2001, 107:3-6), numerous efforts are underway to develop safe inhibitors of NF-kappaB to be used in treatment of both chronic and acute disease situations. Specific inhibitors of NF-kappaB should reduce side effects associated with drugs such as NSAIDs and glucocorticoids and would offer significant potential for the treatment of a variety of human and animal diseases. Specific diseases or syndromes where patients would benefit from NF-kappaB inhibition vary widely, and range from rheumatoid arthritis, atherosclerosis, multiple sclerosis, chronic inflammatory demyelinating polyradiculoneuritis, asthma, inflammatory bowel disease, to *Helicobacter pylori*-associated gastritis and other inflammatory responses, and a variety of drugs that have effects on NF-kappaB activity, such as corticosteroids, sulfasalazine, 5-aminosalicylic acid, aspirin, tepoxalin, leflunomide, curcumin, antioxidants and proteasome inhibitors. These drugs are considered to be non-specific and often only applicable in high concentrations that may end up toxic for the individual treated.

Inactive cytoplasmatic forms of transcription factors can thus be activated by removal of an inhibitor, as is the case with NF-kappaB, or alternatively, by association of two (or more) proteins neither of which is active by itself as in the case of interferon-alpha-stimulated factor (D.E. Levy et al., 1989, *Genes and Development* 3:1362). After interferon-alpha attaches to its cell-surface receptor, one of the proteins is changed within a minute or less, and the two can combine. The active (combined) factor is then translocated



to the cell nucleus to stimulate transcription only of genes with a binding site for the protein. Considering that interferon-alpha is a mediator of responses of the body directed at pathogens and self-antigens, modulating regulation of genes that are under influence of the interferon-alpha-stimulated factor would contribute to the treatment of a variety of human and animal diseases.

Other typical examples of signaling molecules that affect gene expression via cell-surface receptor interaction are polypeptide hormones such as insulin, glucagon, various growth factors such as EGF, VEGF, and so on.

The steroid hormones and their receptors represent one of the best understood cases that affect transcription. Because steroid hormones are soluble in lipid membranes, they can diffuse into cells. They affect transcription by binding to specific intracellular receptors that are site specific DNA-binding molecules. Other examples of signaling molecules that enter the cell and act intra-cellularly are thyroid hormone ( $T_3$ ), vitamin D and retinoic acid, other small lipid-soluble signaling molecules that enter cells and modulate gene expression. The characteristic DNA-binding sites for the receptors for these signaling molecules are also known as response elements.

Another example of a small molecule that is involved in regulation of gene expression is ethylene, a gas that for example induces the expression of genes involved in fruit ripening. Also, small plant hormones, known as auxins and cytokinins regulate plant growth and differentiation directly by regulating gene expression.

Given the critical role of regulatory factors in gene regulation, the development of artificial or synthetic counterparts that could be used in methods to rectify errors in gene expression has been a long standing goal at the interface of chemistry and biology.

## DISCLOSURE OF THE INVENTION

The invention provides a method for modulating expression of a gene in a cell comprising providing the cell with a signaling molecule comprising an oligopeptide or functional analogue or derivative thereof. Such a molecule is herein also called NMPF and referenced by number. Since peptides, and functional analogues and derivatives of relatively short amino acid sequences, are easily synthesized these days, the invention provides a method to modulate gene expression with easily obtainable synthetic compounds such as synthetic peptides or functional analogues or derivatives thereof.

The invention also provides a method for the treatment of an inflammatory condition comprising administering to a subject in need of such treatment a molecule comprising an oligopeptide peptide or functional analogue or derivative thereof the molecule capable of reducing production of NO by a cell, in particular wherein the molecule additionally is capable of modulating translocation and/or activity of a gene transcription factor present in a cell, especially wherein the gene transcription factor comprises a NF-kappaB/Rel protein. Advantageously, the invention provides a method wherein the modulating translocation and/or activity of a gene transcription factor allows modulation of TNF-alpha production by the cell, in particular wherein the TNF-alpha production is reduced. Considering that TNF-alpha production is central to almost all, if not all, inflammatory conditions, reducing TNF-alpha production can greatly alleviate, or mitigate, a great host of inflammatory conditions that are described herein. In particular, the invention provides a method wherein the inflammatory condition comprises an acute inflammatory condition, and it is especially useful to treat anthrax related disease, especially when considering that with anthrax, both NO and TNF-alpha reduction will greatly mitigate the course of disease. Table 6 lists oligopeptides according to the invention that have such modulatory effect. In particular, the invention provides a method of treatment wherein the treatment comprises administering to the subject a pharmaceutical composition comprising an oligopeptide or functional analogue or derivative thereof capable of reducing production of NO by a cell, preferably wherein the composition comprises at least two oligopeptides or functional analogues or derivatives thereof capable of reducing production of NO by a cell, example of such combinations can be selected under guidance of table 6, whereby it suffices to select two, or more, with a desired effect, such as wherein the at least two oligopeptides are selected from the group LQGV, AQGV and VLPALP.

The invention also provides an isolated, preferably synthetic, oligopeptide or functional analogue or derivative thereof or mixture of such oligopeptides or analogues or derivatives capable of reducing production of NO by a cell. Such cell is preferably of a macrophage or DC lineage, considering the central role these cells play in the inflammatory process. The invention also provides a pharmaceutical composition comprising an oligopeptide or functional analogue or derivative according to the invention or comprising at least two oligopeptides or functional analogues or derivatives thereof

capable of reducing production of NO by a cell. Furthermore, the invention provides the use of an oligopeptide or functional analogue or derivative thereof capable of reducing production of NO by a cell for the production of a pharmaceutical composition for the treatment of an inflammatory condition by the reduction of NO production by  
5 macrophages or DC in the subject to be treated.

A functional analogue or derivative of a peptide is defined as an amino acid sequence, or other sequence monomers, which has been altered such that the functional properties of the sequence are essentially the same in kind, not necessarily in amount. An analogue or derivative can be provided in many ways, for instance through conservative  
10 amino acid substitution. Also peptidomimetic compounds can be designed that functionally or structurally resemble the original peptide taken as starting point but that are for example composed of non-naturally occurring amino acids or polyamides. With conservative amino acid substitution, one amino acid residue is substituted with another residue with generally similar properties (size, hydrophobicity, etc), such that the overall functioning is  
15 likely not to be seriously affected. However, it is often much more desirable to improve a specific function. A derivative can also be provided by systematically improving at least one desired property of an amino acid sequence. This can for instance be done by an Ala-scan and/or replacement net mapping method. With these methods, many different peptides are generated, based on an original amino acid sequence but each containing a  
20 substitution of at least one amino acid residue. The amino acid residue may either be replaced by alanine (Ala-scan) or by any other amino acid residue (replacement net mapping). This way, many positional variants of the original amino acid sequence are synthesized. Every positional variant is screened for a specific activity. The generated data are used to design improved peptide derivatives of a certain amino acid sequence.

25 A derivative or analogue can also for instance be generated by substitution of an L-amino acid residue with a D-amino acid residue. This substitution, leading to a peptide which does not naturally occur in nature, can improve a property of an amino acid sequence. It is for example useful to provide a peptide sequence of known activity of all D-amino acids in retro inversion format, thereby allowing for retained activity and increase  
30 half-life values. By generating many positional variants of an original amino acid sequence and screening for a specific activity, improved peptide derivatives comprising such D-amino acids can be designed with further improved characteristics.

A person skilled in the art is well able to generate analogous compounds of an amino acid sequence. This can for instance be done through screening of a peptide library. Such an analogue has essentially the same functional properties of the sequence in kind, not necessarily in amount. Also, peptides or analogues can be circularised (for example by  
5 providing them with (terminal) cysteines, dimerized or multimerized, for example by linkage to lysine or cysteine or other compounds with side-chains that allow linkage or multimerization, brought in tandem- or repeat-configuration, conjugated or otherwise linked to carriers known in the art, if only by a labile link that allows dissociation.

The invention also provides a signaling molecule for modulating expression of a  
10 gene in a cell comprising a small peptide or functional analogue or derivative thereof. Surprisingly, the inventors found that a small peptide acts as a signaling molecule that can modulate signal transduction pathways and gene expression. A functional analogue or derivative of a small peptide that acts as such a signaling molecule for modulating expression of one or more genes in a cell can be identified or obtained by at least one of  
15 various methods for finding such a signaling molecule as provided herein.

For example, one method as provided herein for identifying or obtaining a signaling molecule comprising a peptide or functional derivative or analogue thereof capable of modulating expression of a gene in a cell comprises providing the cell with a peptide or derivative or analogue thereof and determining the activity and/or nuclear  
20 translocation of one or more gene transcription factors. Such activity can be determined in various ways using means and/or methods honed to the specific transcription factor(s) under study. In the detailed description, it is provided to study NF-kappaB/Rel protein translocation and/or activity, but it is of course also easily possible to study translocation and/or activity of any other transcription factor for which such tools are available or can be  
25 designed. One such other transcription factor is for example the interferon-alpha-stimulated factor as discussed above. Other useful transcription factors to study in this context comprise c-Jun, ATF-2, Fos, and their complexes, ELK-1, EGR-1, IRF-1, IRF-3/7, AP-1, NF-AT, C/EBPs, Sp1, CREB, PPARGamma, STAT proteins to name a few. Considering that many proteins are subject to proteolytic breakdown whereby oligopeptide  
30 fragments are generated, many already before the full protein even has exerted a function, it is hereby established that oligopeptide fragments of such proteins (of which a non-extensive list is given in the detailed description, but one can for example think of

MAPKK-2 that can give rise to a peptide MLARRKPVLPALTINP, and subsequently to a peptide comprising MLARRKP or MLAR or VLPALT or VLPAL, but also of nitric oxide synthase that can give rise to peptides FPGC or PGCP, GVLPAVP, LPA, VLPAVP, or PAVP after proper proteolysis) are involved in feedback mechanisms regulating gene expression, likely by the effect of transcription factors on gene expression. In addition, oligopeptide fragments of proteins (of which a non-extensive list is given in the detailed description) can also modulate the activity of extracellular components such as factor XIII (example of oligopeptide fragments obtained from factor XIII are LQGV, LQGVVPRGV, GVVP, VPRGV, PRG, PRGV) or activated protein C (APC) and thereby eventually leading to the modulation of intracellular signal transduction pathways and gene(s) expression.

As said, the invention provides active oligopeptides acting as a signaling molecule. To allow for improved bio-availability of such a signaling molecule (which is useful as a pharmacon, especially when produced artificially) the invention also provides a method for determining whether a small peptide or derivative or analogue thereof can act as a functional signaling molecule according to the invention, the method further comprising determining whether the signaling molecule is membrane-permeable.

The invention for example provides a method a process or method for obtaining information about the capacity or tendency of an oligopeptide, or a modification or derivative thereof, to regulate expression of a gene comprising the steps of:

- a) contacting the oligopeptide, or a modification or derivative thereof, with at least one cell;
- b) determining the presence of at least one gene product in or derived from the cell. It is preferred that the oligopeptide comprises an amino acid sequence corresponding to a fragment of a naturally occurring polypeptide, such as hCG, or MAPKK, or another kinase, be it of plant or animal cell, or of eukaryotic or prokaryotic origin, or a synthase of a regulatory protein in a cell, such as wherein the regulatory protein is a (pro-) inflammatory mediator, such as a cytokine. Several candidate proteins and peptide fragments are listed in the detailed description which are a first choice for such an analysis from the inventors perspective, but the person skilled in the art, and working in a specific field of interest in biotechnology shall

immediately understand which protein to select for such analyses for his or her own purposes related to his or her field.

In particular, it is provided to perform a process according to the invention further comprising a step c) comprising determining the presence of the gene product in or derived from a cell which has not been contacted with the oligopeptide, or a modification or derivative thereof, and determining the ratio of gene product found in step b to gene product found in step c, as can easily be done with the present day genechip technology (see, for example, the detailed description herein) and related methods of expression profiling known in the art.

Another method provided herein for identifying or obtaining information on a signaling molecule, (or for that matter the signaling molecule itself, considering that the next step of synthesizing the molecule, generally being a short peptide, is whole within the art) comprising a peptide or functional derivative or analogue thereof capable of modulating expression of a gene in a cell comprises providing the cell with a peptide or derivative or analogue thereof and determining relative up-regulation and/or down-regulation of at least one gene expressed in the cell. The up-regulation can classically be studied by determining via for example Northern or Western blotting or nucleic acid detection by PCR or immunological detection of proteins whether a cell or cells make more (in the case of up-regulation) or less (in the case of down-regulation) of a gene expression product such as mRNA or protein after the cell or cells have been provided with the peptide or derivative or analogue thereof. Of course, various methods of the invention can be combined to better analyze the functional analogue of the peptide or derivative or analogue under study. Furthermore, relative up-regulation and/or down-regulation of a multitude or clusters of genes expressed in the cell can be easily studied as well, using libraries of positionally or spatially addressable predetermined or known relevant nucleic acid sequences or unique fragments thereof bound to an array or brought in an array format, using for example a nucleic acid library or so-called gene chip expression analysis systems. Lysates of cells or preparations of cytoplasm and/or nuclei of cells that have been provided with the peptide or derivative or analogue under study are then contacted with the library and relative binding of for example mRNA to individual nucleic acids of the library is then determined, as further described herein in the detailed description.

A functional analogue or derivative of a small peptide that can act as a signaling molecule for modulating expression of a gene in a cell can also be identified or obtained by method for identifying or obtaining a signaling molecule comprising an oligopeptide or functional derivative or analogue thereof capable of modulating expression of a gene in a cell comprising providing a peptide or derivative or analogue thereof and determining binding of the peptide or derivative or analogue thereof to a factor related to gene control. Such a factor related to gene control can be any factor related to transcription (either initiation or termination), processing of primary transcripts, stabilization or destabilization of mRNAs, and mRNA translation.

Binding of a peptide or derivative or analogue thereof to such a factor can be determined by various methods known in the art. Classically, peptides or derivatives or analogues can be (radioactively) labeled and binding to the factor can be determined by detection of a labeled peptide-factor complex, such as by electrophoresis, or other separation methods known in the art. However, for determining binding to such factors, array techniques, such as used with peptide libraries, can also be employed, comprising providing a multitude of peptides or derivatives or analogues thereof and determining binding of at least one of the peptides or derivatives or analogues thereof to a factor related to gene control.

In a preferred embodiment, the factor related to gene control comprises a transcription factor, such as a NF-kappaB-Rel protein or another transcription factor desired to be studied. When binding of a functional analogue according to the invention to such factor has been established, it is of course possible to further analyze the analogue by providing a cell with the peptide or derivative or analogue thereof and determining the activity and/or nuclear translocation of a gene transcription factor in the cell, and/or by providing a cell with the peptide or derivative or analogue thereof and determining relative up-regulation and/or down-regulation of at least one gene expressed in the cell.

The invention thus provides a signaling molecule useful in modulating expression of a gene in a cell and/or useful for reducing NO production by a cell and identifiable or obtainable by employing a method according to the invention. Useful examples of such a signaling molecule can be selected from the group of oligopeptides LQG, AQG, LQGV, AQGV, LQGA, VLPALPQVVC, VLPALP, ALPALP, VAPALP, ALPALPQ, VLPAAPQ, VLPALAQ, LAGV, VLAALP, VLAALP, VLPALA, VLPALPQ, VLAALPQ,

VLPALPA, GVLPALP, GVLPALPQ, LQGVLPALPQVVC,  
VVCNYRDVRFESIRLPGCPRGVNPVVSYAVALSCQCAL, RPRCRPINATLAVEK,  
EGCPVCITVNTTICAGYCPT, SKAPPPSLPSPSRLPGPS, SIRLPGCPRGVNPVVS,  
LPGCPRGVNPVVS, LPGC, MTRV, MTR, VVC, QVVC and functional analogues or  
5 derivatives thereof.

A preferred size of a signaling molecule according to the invention is at most 30 –  
40 amino acids, although much smaller molecules, in particular of oligopeptide size, have  
been shown to be particularly effective. Surprisingly, the invention provides here the  
insight that gene expression can be modulated or regulated by small peptides, that are most  
10 likely breakdown products of larger polypeptides such as chorionic gonadotrophin (CG)  
and growth hormones or growth factors such as fibroblast growth factor, EGF, VEGF,  
RNA 3'terminal phosphate cyclase and CAPI8. In principle such regulating peptide  
sequences can be derived from a part of any protein of polypeptide molecule produced by  
prokaryotic and/or eukaryotic cells, and the invention provides the insight that breakdown  
15 products of polypeptides, preferably oligopeptides at about the sizes as provided herein, are  
naturally involved as signaling molecule in modulation of gene expression. In particular, as  
signaling molecule a (synthetic) peptide is provided obtainable or derivable from beta-  
human chorionic gonadotrophin (beta-HCG), preferably from nicked beta-HCG. It was  
thought before that breakdown products of nicked-beta hCG were involved in immuno-  
20 modulation (PCT International Patent Publication WO99/59671) or in the treatment of  
wasting syndrome (PCT International Patent Publication WO97/49721) but a relationship  
with modulation of gene expression was not forwarded in these publications. Of course,  
such an oligopeptide, or functional equivalent or derivative thereof is likely obtainable or  
derivable from other proteins that are subject to breakdown or proteolysis and that are  
25 close to a gene regulatory cascade. Preferably, the peptide signaling molecule is obtained  
from a peptide having at least 10 amino acids such as a peptide having an amino acid  
sequence MTRVLQGVLPALPQVVC, SIRLPGCPRGVNPVVS,  
VVCNYRDVRFESIRLPGCPRGVNPVVSYAVALSCQCAL,  
RPRCRPINATLAVEKEGCPVCITVNTTICAGYCPT,  
30 CALCRRSTTDCGGPKDHPLTC, SKAPPPSLPSPSRLPGPS,  
CRRSTTDCGGPKDHPLTC, TCDDPRFQDSSSSKAPPPSLPSPSRLPGPSDTPILPQ or  
functional fragment (e.g., a breakdown product) or functional analogue thereof. Functional



analogue herein relates to the signaling molecular effect or activity as for example can be measured by measuring nuclear translocation of a relevant transcription factor, such as NF-kappaB in an NF-kappaB assay, or AP-1 in an AP-1 assay, or by another method as provided herein. Fragments can be somewhat (*i.e.*, 1 or 2 amino acids) smaller or larger on one or both sides, while still providing functional activity.

Not wishing to be bound by theory, it is postulated herein that an unexpected mode of gene regulation has been uncovered. Polypeptides, such as endogenous CG, EGF etc, but also polypeptides of pathogens such as viral, bacterial or protozoal polypeptides, are subject to breakdown into distinct oligopeptides, for example by intracellular proteolysis.

Distinct proteolytic enzymes are widely available in the cell, for example in eukaryotes in the lysosomal or proteasomal system. Some of the resulting breakdown products are oligopeptides of 3 to 15, preferably 4 to 9, most preferably 4 to 6 amino acids long that are surprisingly not without any function or effect to the cell, but as demonstrated herein may be involved, possibly via a feedback mechanism in the case of breakdown of endogenous polypeptides, as signaling molecules in the regulation of gene expression, as demonstrated herein by the regulation of the activity or translocation of a gene transcription factor such as NF-kappaB by for example peptide LQGV, VLPALPQVVC, LQGVLPALPQ, LQG, GVLPALPQ, VLPALP, VLPALPQ, GVLPALP, VVC, MTRV, MTR. Synthetic versions of these oligopeptides as described above, and functional analogues or derivatives of these breakdown products are herein provided to modulate gene expression in a cell and be used in methods to rectify errors in gene expression or the treatment of disease. Oligopeptides such as LQG, AQG, LQGV, AQGV, LQGA, VLPALP, ALPALP, VAPALP, ALPALPQ, VLPAAPQ, VLPALAQ, LAGV, VLAALP, VLAALP, VLPALA, VLPALPQ, VLAALPQ, VLPALPA, GVLPALP, GVLPALPQ, LQGVLPALPQVVC, SIRLPGCPRGVNPVVS, SKAPPPSLPSPSRLPGPS, LPGCPRGVNPVVS, LPGC, MTRV, MTR, VVC, or functional analogues or derivatives (including break down products) of the longer sequences thereof, are particularly effective.

By using the insight as expressed herein, in a preferred embodiment, the invention provides a method for modulating expression of a gene in a cell comprising providing the cell with a signaling molecule comprising an oligopeptide or functional analogue or derivative thereof wherein the signaling molecule is membrane-permeable in that it enters the cell. Most small peptides as described herein have already an inherent propensity to

become intracellularly involved, but signaling molecules as provided herein can also be provided with additional peptide sequences, such as arginine or lysine rich stretches of amino acids, that allow for improved internalization across a lipid bilayer membrane, and may possibly be cleaved off later by internal proteolytic activity.

5           In a preferred embodiment, the invention provides a method for modulating expression of a gene in a cell comprising providing the cell with a signaling molecule comprising a small peptide (amino acid sequence) or functional analogue or derivative thereof, wherein the signaling molecule modulates NF-kappaB/Rel protein conversion or translocation. As said, NF-κB was originally identified as a gene transcription factor that  
10   bound to an enhancer element in the gene for the Igk light chain and was believed to be B cell-specific. However, subsequent studies revealed that NF-kappaB/Rel proteins are ubiquitously expressed and play a central role as transcription factor in regulating the expression of many genes, particularly those involved in immune, inflammatory, developmental and apoptotic processes. NF-κB related gene transcription factors can be  
15   activated by different stimuli such as microbial products, proinflammatory cytokines, T- and B-cell mitogens, and physical and chemical stresses. NF-κB in turn regulates the inducible expression of many cytokines, chemokines, adhesion molecules, acute phase proteins, and antimicrobial peptides.

          NF-κB represents a group of structurally related and evolutionarily conserved gene  
20   transcription factors. So far, five mammalian NF-κB proteins named Rel (c-Rel), RelA (p65), RelB, NF-κB (p50 and its precursor p105), and NF-κB (p52 and its precursor p100) have been described. NF-κB proteins can exist as homo- or heterodimers, and although most NF-κB dimers are activators of transcription, the p50/p50 and p52/p52 homodimers often repress the transcription of their target genes. In *Drosophila*, three NF-κB homologs  
25   named Dorsal, Dif, and Relish have been identified and characterized. Structurally, all NF-κB/Rel proteins share a highly conserved NH<sub>2</sub>-terminal Rel homology domain (RHD) that is responsible for DNA binding, dimerization, and association with inhibitory proteins known as IκBs. In resting cells, NF-κB/Rel dimers are bound to IκBs and retained in an inactive form in the cytoplasm. Like NF-κB, IκBs are also members of a multigene family  
30   containing seven known mammalian members including IκBα, IκBβ, IκBγ, IκBε, Bcl-3,

the precursor Rel-proteins, p100, and p105, and one *Drosophila* IκB named Cactus. The IκB family is characterized by the presence of multiple copies of ankyrin repeats, which are protein-protein interaction motifs that interact with NF-κB via the RHD. Upon appropriate stimulation, IκB is phosphorylated by IκB kinases (IKKs), polyubiquitinated by an ubiquitin ligase complex, and degraded by the 26S proteasome. Consequently, NF-κB is released and translocates into the nucleus to initiate gene expression.

NF-κB related transcription factors regulate the expression of a wide variety of genes that play critical roles in innate immune responses. Such NF-κB target genes include those encoding cytokines (*e.g.*, IL-1, IL-2, IL-6, IL-12, TNF-α, LTα, LTβ, and GM-CSF), adhesion molecules (*e.g.*, ICAM, VCAM, endothelial leukocyte adhesion molecule [ELAM]), acute phase proteins (*e.g.*, SAA), and inducible enzymes (*e.g.*, iNOS and COX-2). In addition, it has been demonstrated recently that several evolutionary conserved antimicrobial peptides, for example, β-defensins, are also regulated by NF-κB, a situation similar to *Drosophila*. Besides regulating the expression of molecules involved in innate immunity, NF-κB also plays a role in the expression of molecules important for adaptive immunity, such as MHC proteins, and the expression of critical cytokines such as IL-2, IL-12 and IFN-γ. Finally, NF-κB plays an important role in the overall immune response by affecting the expression of genes that are critical for regulating the apoptotic process, such as c-IAP-1 and c-IAP-2, Fas ligand, c-myc, p53, and cyclin D1.

Under normal conditions, NF-kappaB is rapidly activated upon microbial and viral invasion, and this activation usually correlates with resistance of the host to infection. However, persistent activation of NF-kappaB may lead to the production of excessive amounts of pro-inflammatory mediators such as IL-12 and TNF-alpha, resulting in tissue damage, as in insulin dependent diabetes mellitus, atherosclerosis, Crohn's disease, organ failure, and even death of the host, as in bacterial infection-induced septic shock. It is interesting to note that in order to survive in the host, certain pathogens, such as *Bordetella*, *Yersinia*, *Toxoplasma gondii* and African Swine Fever Virus have evolved mechanisms to counteract or escape the host system by inhibiting NF-kappaB activation. On the other hand, some viruses, including HIV-1, CMV and SV-40 take advantage of NF-kappaB as a host factor that is activated at sites of infection.

Furthermore, the invention provides a method to explore alterations in gene expression in antigen presenting cells such as dendritic cells in response to microbial exposure by analyzing a gene-expression profile of dendritic cells in response to micro organisms such as for example bacteria such as *Escherichia coli*, or other pathogenic bacteria, fungi or yeasts such as *Candida albicans*, viruses such as influenza virus and the effect of (simultaneous) treatment of these diseases with a signaling molecule according to the invention. For example, human monocyte-derived dendritic cells are cultured with one or more pathogens for 1-36 hours, and gene expression is analyzed using an oligonucleotide array representing a (be it large or small) set of genes. When the pathogens regulate the expression of a core set of a distinct number genes, these genes may be classified according to their kinetics of expression and function. Generally, within 4 hours of pathogen exposure, genes associated with pathogen recognition and phagocytosis will be down-regulated, whereas genes for antigen processing and presentation are up-regulated- 8 hours post-exposure. Treatment of such dendritic cells with a signaling molecule according to the invention (be it simultaneous or before or after the treatment of the cells with the pathogen) allows studying the effect of a signaling molecule according to the invention has on the effect a pathogen has on an antigen presenting cell.

In short, the invention surprisingly provides a signaling molecule capable of modulating expression of a gene in a cell, the molecule being a short peptide, preferably of at most 30 amino acids long or a functional analogue or derivative thereof. In a much preferred embodiment, the peptide is an oligopeptide of from about 3 to at about 15 amino acids long, preferably 4 to 12, more preferably 4 to 9, most preferably 4 to 6 amino acids long, or a functional analogue or derivative thereof. Of course, such signaling molecule can be longer, for example by extending it (N- and/or C-terminally, with more amino acids or other side groups, which can for example be (enzymatically) cleaved off when the molecule enters the place of final destination. Such extension may even be preferable to prevent the signaling molecule from becoming active in an untimely fashion; however, the core or active fragment of the molecule comprises the aforementioned oligopeptide or analogue or derivative thereof.

In particular, the invention provides a modulator of NF-kappaB/Rel protein activation comprising a signaling molecule according to the invention. Such modulators are widely searched after these days. Furthermore, the invention provides use of a

signaling molecule according to the invention for the production of a pharmaceutical composition for the modulation of gene expression.

Also, the invention provides a method for the treatment of bone disease such as osteoporosis comprising administering to a subject in need of such treatment a molecule comprising an oligopeptide peptide or functional analogue thereof the molecule capable of modulating production of NO and/or TNF-alpha by a cell. Such a method of treatment is particularly useful in post-menopausal women that do not longer experience the benefits of being provided with a natural source of several of the signaling molecules as provided herein, hCG and its breakdown products. Furthermore, the invention provides method for the treatment of an inflammatory condition associated with TNF-alpha activity of fibroblasts, such as seen with chronic arthritis or synovitis comprising administering to a subject in need of such treatment a molecule comprising an oligopeptide peptide or functional analogue thereof wherein the molecule is capable of modulating translocation and/or activity of a gene transcription factor present in a cell, in particular of the NF-kappaB factor. Such a treatment can be achieved by systemic administration of a signaling molecule according to the invention, but local administration in joints, bursae or tendon sheaths is provided as well. The molecule can be selected from table 6 or identified in a method according to the invention. It is preferred when the treatment comprises administering to the subject a pharmaceutical composition comprising an oligopeptide or functional analogue thereof also capable of reducing production of NO by a cell, for example wherein said composition comprises at least two oligopeptides or functional analogues thereof each capable of reducing production of NO and/or TNF-alpha by a cell, in particular wherein the at least two oligopeptides are selected from the group LQGV, AQGV and VLPALP.

Furthermore, the invention provides use of an oligopeptide or functional analogue thereof capable of reducing production of NO and/or TNF-alpha by a cell for the production of a pharmaceutical composition for the treatment of an inflammatory condition or a post-meno-pausal condition, or a bone disease such as osteoporosis, or for the induction of weight loss. The term pharmaceutical composition as used herein is intended to cover both the active signaling molecule alone or a composition containing the signaling molecule together with a pharmaceutically acceptable carrier, diluent or excipient. Acceptable diluents of an oligopeptide as described herein in the detailed description are

for example physiological salt solutions or phosphate buffered salt solutions. In one embodiment of the present invention, a signal molecule is administered in an effective concentration to an animal or human systemically, for instance, by intravenous, intramuscular or intraperitoneal administration. Another way of administration comprises perfusion of organs or tissue, be it *in vivo* or *ex vivo*, with a perfusion fluid comprising a signal molecule according to the invention. Topical administration, e.g., in ointments or sprays, may also apply, for example, in inflammations of the skin or mucosal surfaces of for example mouth, nose and/or genitals. Local administration can occur in joints, bursae, tendon sheaths, in or around the spinal cord at locations where nerve bundles branch off, at the location of hernias, in or around infarcted areas in brain or heart, etc. The administration may be done as a single dose, as a discontinuous sequence of various doses, or continuously for a period of time sufficient to permit substantial modulation of gene expression. In the case of a continuous administration, the duration of the administration may vary depending upon a number of factors that would readily be appreciated by those skilled in the art.

The administration dose of the active molecule may be varied over a fairly broad range. The concentrations of an active molecule that can be administered would be limited by efficacy at the lower end and the solubility of the compound at the upper end. The optimal dose or doses for a particular patient should and can be determined by the physician or medical specialist involved, taking into consideration well known relevant factors such as the condition, weight and age of the patient, etc.

The active molecule may be administered directly in a suitable vehicle, such as for example, phosphate-buffered saline (PBS) or as solutions in alcohol or DMSO. Pursuant to preferred embodiments of the present invention, however, the active molecule is administered through a single dose delivery using a drug-delivery system, such as a sustained-release delivery system, which enables the maintenance of the required concentrations of the active molecule for a period of time sufficient for adequate modulation of gene expression. A suitable drug-delivery system would be pharmacologically inactive or at least tolerable. It should preferably not be immunogenic nor cause inflammatory reactions, and should permit release of the active molecule so as to maintain effective levels thereof over the desired time period. A large variety of alternatives are known in the art as suitable for purposes of sustained release and are

contemplated as within the scope of the present invention. Suitable delivery vehicles include, but are not limited to, the following: microcapsules or microspheres; liposomes and other lipid-based release systems; viscous instillates; absorbable and/or biodegradable mechanical barriers and implants; and polymeric delivery materials, such as polyethylene oxide/polypropylene oxide block copolymers, polyesters, cross-linked polyvinylalcohols, polyanhydrides, polymethacrylate and polymethacrylamide hydrogels, anionic carbohydrate polymers, etc. Useful delivery systems are well known in the art.

A highly suitable formulation to achieve the active molecule release comprises injectable microcapsules or microspheres made from a biodegradable polymer, such as poly (dl-lactide), poly (dl-lactide-co-glycolide), polycaprolactone, polyglycolide, polylactic acid-co-glycolide, poly (hydroxybutyric acid), polyesters or polyacetals. Injectable systems comprising microcapsules or microspheres having a diameter of about 50 to about 500 micrometers offer advantages over other delivery systems. For example, they generally use less active molecule and may be administered by paramedical personnel. Moreover, such systems are inherently flexible in the design of the duration and rate of separate drug release by selection of microcapsule or microsphere size, drug loading and dosage administered. Further, they can be successfully sterilized by gamma irradiation.

The design, preparation and use of microcapsules and microspheres are well within the reach of persons skilled in the art and detailed information concerning these points is available in the literature. Biodegradable polymers (such as lactide, glycolide and caprolactone polymers) may also be used in formulations other than microcapsules and microspheres; for example, pre-made films and spray-on films of these polymers containing the active molecule would be suitable for use in accordance with the present invention. Fibers or filaments comprising the active molecule are also contemplated as within the scope of the present invention.

Another highly suitable formulation for a single-dose delivery of the active molecule in accordance with the present invention involves liposomes. The encapsulation of an active molecule in liposomes or multilamellar vesicles is a well known technique for targeted drug delivery and prolonged drug residence. The preparation and use of drug-loaded liposomes is well within the reach of persons skilled in the art and well documented in the literature.

Yet another suitable approach for single dose delivery of an active molecule in accordance with the present invention involves the use of viscous instillates. In this technique, high molecular weight carriers are used in admixture with the active molecule, giving rise to a structure that produces a solution with high viscosity. Suitable high molecular weight carriers include, but are not limited to, the following: dextrans and cyclodextrans; hydrogels; (cross-linked) viscous materials, including (cross-linked) viscoelastics; carboxymethylcellulose; hyaluronic acid; and chondroitin sulfate. The preparation and use of drug-loaded viscous instillates is well known to persons skilled in the art.

Pursuant to yet another approach, the active molecule may be administered in combination with absorbable mechanical barriers such oxidised regenerated cellulose. The active molecule may be covalent or non-covalently (e.g., ionically) bound to such a barrier, or it may simply be dispersed therein. A pharmaceutical composition as provided herein is particularly useful for the modulation of gene expression by inhibiting NF-kappaB/Rel protein activation.

NF-kappaB/Rel proteins are a group of structurally related and evolutionarily conserved proteins (Rel). Well known are c-Rel, RelA (p65), RelB, NF-kappaB1 (p50 and its precursor p105), and NF-kappaB2 (p52 and its precursor p100). Most NF-kappaB dimers are activators of transcription; p50/p50 and p52/p52 homodimers repress the transcription of their target genes. All NF-kappaB/Rel proteins share a highly conserved NH2-terminal Rel homology domain (RHD). RHD is responsible for DNA binding, dimerization, association with inhibitory proteins known as IkappaBs. In resting cells, NF-kappaB/Rel dimers are bound to IkappaBs and retained in an inactive form in the cytoplasm. IkappaBs are members of a multigene family (IkappaBalpha, IkappaBbeta, IkappaBgamma, IkappaBepsilon, Bcl-3, the precursor Rel-proteins, p100 and p105. Presence of multiple copies of ankyrin repeats to interact with NF-kappaB via the RHD (protein-protein interaction. Upon appropriate stimulation, IkappaB is phosphorylated by IkappaB Kinase (IKKs), polyubiquitinated by ubiquitin ligase complex, and degraded by the 26S proteasome. NF-kappaB is released and translocates into nucleus to initiate gene expression.

NF-kappaB regulation of gene expression includes innate immune responses: such as regulated by cytokines IL-1, IL-2, IL-6, IL-12, TNF-alpha, LT-alpha, LT-beta,



GM-CSF; expression of adhesion molecules (ICAM, VCAM, endothelial leukocyte adhesion molecule [ELAM]), acute phase proteins (SAA), inducible enzymes (iNOS and COX-2) and antimicrobial peptides (beta-defensins). For adaptive immunity MHC proteins, IL-2, IL-12 and IFN-alpha are regulated by NF-kappaB. Regulation of overall immune response includes the regulation of genes critical for regulation of apoptosis (c-IAP-1 and c-IAP-2, Fas Ligand, c-myc, p53 and cyclin D1.

Considering that NF-kappaB and related transcription factors are cardinal pro-inflammatory transcription factors, and considering that the invention provides a signaling molecule, such as an oligopeptide and functional analogues or derivatives thereof that are capable of inhibiting NF-kappaB and likely also other pro-inflammatory transcription factors, herein also called NF-kappaB inhibitors, the invention provides a method for modulating NF-kappaB activated gene expression, in particular for inhibiting the expression and thus inhibiting a central pro-inflammatory pathway.

The consequence of this potency to inhibit this pro-inflammatory pathway is wide and far reaching. The invention for example provides a method to mitigate or treat inflammatory airway disease such as asthma. Generally, asthma patients show persistent activation of NF-kappaB of cells lining the respiratory tract. Providing these patients, for example by aerosol application, with a signaling molecule according to the invention, such as LQGV or AQGV or MTRV or functional analogue or derivative thereof, will alleviate the inflammatory airway response of these individuals, by inhibiting NF-kappaB activation of the cells. Such compositions can advantageously be made with signaling molecules that are taken up in liposomes.

As said, inflammation involves the sequential activation of signaling pathways leading to the production of both pro- and anti-inflammatory mediators. Considering that much attention has focused on pro-inflammatory pathways that initiate inflammation, relatively little is known about the mechanisms that switch off inflammation and resolve the inflammatory response. The transcription factor NF-kB is thought to have a central role in the induction of pro-inflammatory gene expression and has attracted interest as a new target for the treatment of inflammatory disease. However NF-kB activation of leukocytes recruited during the onset of inflammation is also associated with pro-inflammatory gene expression, whereas such activation during the resolution of inflammation is associated with the expression of anti-inflammatory genes and the induction of apoptosis. Inhibition

of NF-kB during the resolution of inflammation protracts the inflammatory response and prevents apoptosis. This shows that NF-kB has an anti-inflammatory role *in vivo* involving the regulation of inflammatory resolution. The invention provides a tool to modulate the inflammation at the end phase, a signaling molecule or modulator as provided herein  
5 allows the modulation of the NF-kappaB pathway at different stages of the inflammatory response in vivo, in a particular embodiment, the invention provides a modulator of NF-kappaB for use in the resolution of inflammation, for example through the regulation of leukocyte apoptosis. Useful oligopeptides can be found among those that accelerate shock.

The invention also provides a method to mitigate or treat neonatal lung disease, also  
10 called chronic lung disease of prematurity, a condition often seen with premature children who develop a prolonged pulmonary inflammation or bronchopulmonary dysplasia. Treating such premature children with an NF-kappaB inhibitor, such as oligopeptide LQGV, or functional analogue or derivative thereof, as provided herein allows such lung conditions to be prevented or ameliorated as well.

Recent advances in bone biology provide insight into the pathogenesis of bone  
15 diseases. The invention also provides a method of treatment of a post-menopausal condition such as osteoporosis comprising modulation and inhibition of osteoclast differentiation and inhibiting TNF-alpha induced apoptosis of osteoblasts, thereby limiting the dissolve of bone structures, otherwise so prominent in post-menopausal women that  
20 have no longer a natural source of hCG and thus lack the modulatory effect of the signal molecules that are derived of hCG as shown herein. The invention thus also provides a method of treatment of a bone disease, such as osteoporosis (which is often, but not exclusively seen with post-meno-pausal women) Furthermore, NO and TNF-alpha modulators as provided herein inhibit the inflammatory response and bone loss in  
25 periodontitis. Furthermore, considering that there is a correlation between TNF-alpha activity, and severity of clinical manifestations in ankylosing spondylitis, the invention provides the treatment of spondylitis by use of a signaling molecule as provided herein.

Furthermore, considering that an important pathogenic component in the development of insulin-dependent diabetes mellitus (type 1) comprises over-activation of  
30 the NF-kappaB pathway as seen in dendritic cells, treatment with an NF-kappaB inhibitor according to the invention will lead to reduced symptoms of diabetes, or at least to a prolonged time to onset of the disease. Particularly effective oligopeptide signaling

molecules according to the invention in this context are GVLPALPQ, LQGV MTRV, VLPALPQVVC, VLPALP, VLPALPQ, LPGCPRGVNPVVS, LPGC, VVCNYRDVRFESIRLPGCPRGVNPVVS, YAAVALSCQCAL, CPRGVNPVVS , which were shown herein to postpone onset of diabetes in a Non-obese Diabetic Mouse (NOD).

- 5 Another approach to treatment of diabetes, in particular insulin –independent diabetes (type 2) comprises inhibition of the PPARgamma cascade with an oligopeptide signaling molecule or functional analogue or derivative thereof.

Other use that is provided relates to a method for combating or treating auto-immune disease. A non-limiting list of immune diseases includes:

- 10 Hashimoto's thyroiditis, primary myxedema thyrotoxicosis, pernicious anemia, autoimmune atrophic gastritis, Addison's disease, premature menopause, insulin-dependent diabetes mellitus, stiff-man syndrome, Goodpasture's syndrome, myasthenia gravis, male infertility, pemphigus vulgaris, pemphigoid, sympathetic ophthalmia, phacogenic uveitis, multiple sclerosis, autoimmune haemolytic anemia, idiopathic
- 15 thrombocytopenic purpura, idiopathic leucopenia, primary biliary cirrhosis, active chronic hepatitis, autoimmune deafness, cryptogenic cirrhosis, ulcerative colitis, Sjögren's syndrome, rheumatoid arthritis, dermatomyositis, polymyositis, scleroderma, mixed connective tissue disease, discoid lupus erythematosus, and systemic lupus erythematosus.

- Other use that is provided relates to a method for combating or treating infections
- 20 caused by micro-organisms, in particular those infections that are caused by micro-organisms that activate the NF-kappaB pathway during infections.

- Such micro-organisms are manifold, including bacteria, viruses, fungi, protozoa, but other pathogens (e.g., worms) can have the same effect. Activation of the NFkappaB pathway by a microbial infection in general occurs via activation of the Toll-like receptor
- 25 pathway. The invention provides a method to modulate and in particular to inhibit parts of gene expression that are related to the inflammatory responses of an organism that are generally activated through one of the Toll-like receptor pathways.

- Toll-like receptor-mediated NF-kappa-B activation is central in recognition of pathogens by a host. Such recognition of pathogens generally occurs through germline-
- 30 encoded molecules, the so-called pattern recognition receptors (PRRs). These PRRs recognize widespread pathogen associated molecular patterns (PAMPs). The pattern recognition receptors are expressed as either membrane-bound or soluble proteins. They

include CD14, beta2-integrins (CD11/CD18), C-type lectins, macrophage scavenger receptors, complement receptors (CR1/CD35, CR2/CD21) and Toll-like receptors (TLRs). TLRs are distinguished from other PRRs by their ability to recognize and discriminate between different classes of pathogens. TLRs represent a family of transmembrane proteins that have an extracellular domain comprising multiple copies of leucine rich repeats (LRRs) and a cytoplasmic: Toll/IL-1R (TIR) motif that has significant homology to the intracellular signaling domain of the type I IL-1 receptor (IL-1RI). Therefore, TLRs are thought to belong to the IL-1R superfamily.

Pathogen associated molecular patterns (PAMPS) are not expressed by hosts but are components of the pathogenic micro-organism. Such PAMPS comprise bacterial cell wall components such as lipopolysaccharides (LPS), lipoproteins (BLP), peptidoglycans (PGN), lipoarabinomannan (LAM), lipoteichoic acid (LTA), DNA containing unmethylated CpG motifs, yeast and fungal cell walls mannans and beta-glucans, double stranded RNA, several unique glycosylated proteins and lipids of protozoa, and so on.

Recognition of these PAMPS foremost provides for differential recognition of pathogens by TLRs. For example, TLR2 is generally activated in response to BLPs, PGNs of gram-positive bacteria, LAM of mycobacteria, and mannans of yeasts, whereas TLR4 is often activated by LPS of gram-negative bacteria and LTA of gram-negative bacteria; also a secreted small molecule MD-2 can account for TLR4 signaling.

Several oligopeptides capable of modulating gene expression according to the invention have earlier been tested, both *ex vivo* as *in vivo*, and in small animals but a relationship with modulation of gene expression was not brought forward. A beneficial effect of these oligopeptides on LPS induced sepsis in mice, namely the inhibition of the effect of the sepsis, was observed. Immunomodulatory effects with these oligopeptides have been observed *in vitro* and *in ex vivo* such as in T-cell assays showing the inhibition of pathological Th1 immune responses, suppression of inflammatory cytokines (MIF), increase in production of anti-inflammatory cytokines (IL-10, TGF-beta) and immunomodulatory effects on antigen presenting cells (APC) like dendritic cells, monocytes and macrophages.

Now that the insight has been provided that distinct synthetic oligopeptides or functional analogues or derivatives thereof, for example those that resemble breakdown products which can be derived by proteolysis from endogenous proteins such as hCG, can

be used to modulate gene expression, for example, by NF-kappaB inhibition, such oligopeptides find much wider application. Release of active NF-kappaB in cells is now known to occur after a variety of stimuli including treating cells with bacterial lipopolysaccharide (LPS) and the interaction with a Toll-like receptor (see for example  
5 Guha and Mackman, *Cell. Sign.* 2001, 13:85-94). In particular, LPS stimulation of dendritic cells, monocytes and macrophages induces many genes that are under the influence of activation by transcription factors such as NF-kappaB, p50, EGR-1, IRF-1 and others that can be modulated by a signaling molecule according to the invention. Considering that LPS induction of EGR-1 is required for maximal induction of TNF-alpha,  
10 it is foreseen that inhibition of EGR-1 considerably reduces the effects of sepsis seen after LPS activation. Now knowing the gene modulatory effect of the signaling molecules such as oligopeptides as provided herein allows for rational design of signal molecule mixtures that better alleviate the symptoms seen with sepsis. One such mixture, a 1:1:1 mixture of LQGV, AQGV and VLPALP was administered to primates in a gram-negative induced  
15 rhesus monkey sepsis model for prevention of septic shock and found to be effective in this primate model. Accordingly, the invention provides a pharmaceutical composition for the treatment of sepsis in a primate, and a method for the treatment of sepsis in a primate comprising subjecting the primate to a signaling molecule according to the invention, preferably to a mixture of such signaling molecules. Administration of such a signaling  
20 molecule or mixture preferably occurs systematically, e.g., by intravenous or intraperitoneal administration. In a further embodiment, such treatment also comprises the use of for example an antibiotic, however, only when such use is not contra-indicated because of the risk of generating further toxin loads because of lysis of the bacteria subject to the action of those antibiotics in an individual thus treated.

25 Other use that is contemplated relates to a method for combating or treating viral infections, in particular those infections that are caused by viruses that activate the NF-kappaB pathway during infections. Such virus infections are manifold, classical examples are hepatitis B virus induced cell transformation by persistent activation of NF-kappaB. Use of a signaling molecule according to the invention is herein provided to counter or  
30 prevent this cell transformation.

Other disease where persistent NF-kappaB activation is advantageously inhibited by a signaling molecule according to the invention is a transplantation-related disease such

as transplantation-related immune responses, graft-versus-host-disease, in particular with bone-marrow transplants, acute or chronic xeno-transplant rejection, and post-transfusion thrombocytopenia.

Other case where persistent NF-kappaB activation is advantageously inhibited by a signaling molecule according to the invention is found in the prevention or mitigation of ischemia-related tissue damage seen after infarcts, seen for example in vivo in brain or heart, or ex vivo in organs or tissue that is being prepared or stored in preparation of further use as a transplant. Ischemia related tissue damage can now be mitigated by perfusing the (pre)ischemic area with a signaling molecule according to the invention that inhibits NF-kappaB activation. Example of conditions where ischemia (also called underperfusion) plays a role are for example eclampsia which can be ascribed to focal cerebral ischemia resulting from vasoconstriction, consistent with the evidence of changes detected by new cerebral imaging techniques. The liver dysfunction intrinsic to the HELLP (hemolysis, elevated liver enzymes, and low platelet count) syndrome could also be attributed to the effects of acute underperfusion. Other conditions of ischemia are seen after coronary occlusion leading to irreversible myocardial damage produced by prolonged episodes of coronary artery occlusion and reperfusion in vivo, which has already been discussed in International Patent Application PCT/NL01/00259 as well.

Now that the insight has been provided that distinct synthetic oligopeptides, for example those that resemble breakdown products which can be derived by proteolysis from endogenous proteins such as hCG, can be used to modulate gene expression, for example by NF-kappaB inhibition, the oligopeptides find much wider application. For example, the invention provides a method for perfusing a transplant with a perfusing fluid comprising at least one signaling molecule according to the invention; ischemic or pre-implantation damage due to activation of NF-kappaB in the transplant can than greatly be diminished, allowing a wider use of the transplants.

The invention provides a signaling molecule useful in modulating expression of a gene in a cell. Several examples of the use of such a signaling molecule for the production of a pharmaceutical composition for the treatment of medical or veterinary conditions are herewith given. In one embodiment, the invention provides such use in the treatment of an immune-mediated-disorder, in particular of those cases whereby a central role of NF-kappaB/Rel proteins in the immune response is found. However as said,

modulating gene expression via modulating activity of other transcription factors, such as AP-1 or PPARgamma , and others is also provided, now that the gene modulating role of signaling molecules such as the oligopeptides or analogues or derivatives thereof is understood. As also said, now knowing that oligopeptides, likely breakdown products, play such a central role in modulation of gene expression, the invention provides straightforward ways for identifying further gene expression modulating oligopeptides, and provides synthetic versions of these , and analogues and derivatives thereof for use in a wide variety of disorders and for use in the preparation of a wide variety of pharmaceutical compositions. Examples of such treatment and useful pharmaceutical compositions are for example found in relation to conditions wherein the immune-mediated disorder comprises chronic inflammation, such as diabetes, multiple sclerosis or acute or chronic transplant rejection, in particular in those cases whereby antigen presenting cells (APC's) or dendritic cells (DCs) are enhanced by (overactive) and persistent NF-kappaB expression or wherein the immune-mediated disorder comprises acute inflammation, such as septic or anaphylactic shock or acute transplant rejection. Other immune-mediated disorders that can be treated with a pharmaceutical composition comprising a signaling molecule according to the invention comprise auto-immune disease, such as systemic lupus erythematosus or rheumatoid arthritis (in particular by inhibiting IL-8 and/or IL-15 production by inhibiting NF-kappaB activity on the expression of these genes), allergy, such as asthma or parasitic disease, overly strong immune responses directed against an infectious agent, such as a virus or bacterium (in particular responses that include rapid hemorrhagic disease caused by infection with organisms such as *Yersinia pestis*, Ebola-virus, *Staphylococcus aureus* (e.g., in cases of tampon-disease), bacterial (such as meningococcal) or viral meningitis and/or encephalitis, and other life-threatening conditions). Such overly strong responses are seen with for example pre-eclampsia, recurrent spontaneous abortions (RSA) or preterm parturition or other pregnancy related disorders. Especially with forms of eclampsia/pre-eclampsia that are associated with genetically programmed increased production of tumor-growth factor beta-1, treatment according to the invention is recommended. Also, in situations where RSA is likely attributable to increased IL-10 levels during pregnancy, or to increased TNF-alpha activity, for example due to the presence of an unfavorable allele, in particular of a G to A polymorphism in the promoter of the gene encoding TNF-alpha, treatment with a

pharmaceutical composition as provided herein is recommended. Treatment directed at such pregnancy-related immune disorders is herein also provided by inhibiting NF-kappaB activity directed at activating natural killer (NK) cell activity. Also, LPS induced reduced fertility, or abortions, seen in pregnant sows, can be reduced by applying a signaling molecule or method as provided herein.

Such use in treatment of an immune-mediated disorder preferably comprises regulating relative ratios and/or cytokine activity of lymphocyte, dendritic or antigen presenting cell subset-populations in a treated individual, in particular wherein the subset populations comprise Th1 or Th2, or DC1 or DC2 cells. Other embodiments of the invention comprise use of a signaling molecule according to the invention for the manufacture of a medicament for modulating a cardiovascular or circulatory disorder, such as coronary arterial occlusion and also in a pregnancy related cardiovascular or circulatory disorder.

Furthermore, the invention provides a pharmaceutical composition for modulating a cardiovascular or circulatory disorder, in particular a pregnancy related cardiovascular or circulatory disorder, comprising a signaling molecule according to the invention or mixtures thereof. Such a composition finds its use in a method for modulating a cardiovascular or circulatory disorder, in particular a pregnancy related cardiovascular or circulatory disorder, comprising subjecting an animal (in particular a mammal) to treatment with at least one signaling molecule according to the invention. Non-pregnancy related disorders, which are for example related to hypercholesterolemia, are susceptible to treatment with a signaling molecule according to the invention as well. For example, apolipoprotein E (apo E) deficiency is associated with a series of pathological conditions including dyslipidemia, atherosclerosis, Alzheimer's disease, increase body weight and shorter life span. Inheritance of different alleles of the POLYMORPHIC *apoE* gene is responsible for 10% of the variation in plasma cholesterol in most populations. Individuals HOMOZYGOUS for one variant, *apoE2*, can develop type III dysbetalipoproteinemia if an additional genetic or environmental factor is present. Some much rarer alleles of *apoE* produce dominant expression of this disorder in heterozygous individuals. ApoE, is a ligand for the LDL receptor and its effects on plasma cholesterol are mediated by differences in the affinity of the LDL receptor for lipoproteins carrying variant apoE proteins. The factors that regulate *apoE* gene transcription have been investigated



extensively by the expression of gene constructs in transgenic mice and involve complex interactions between factors that bind elements in the 5' promoter region, in the first intron and in 3' regions many kilobases distant from the structural gene. Deletion of the apo E gene is associated with changes in lipoprotein metabolism [plasma total cholesterol), HDL  
5 cholesterol, HDL/TC, and HDL/LDL ratios, esterification rate in apo B-depleted plasma, plasma triglyceride, hepatic HMG-CoA reductase activity, hepatic cholesterol content, decreased plasma homocyst(e)ine and glucose levels, and severe atherosclerosis and cutaneous xanthomatosis. The invention provides a method and a signaling molecule for the treatment of conditions that are associated with dysfunctional LDL receptors such as  
10 ApoE and other members of the apolipoprotein family. In particular, use of a signaling molecule comprising GVLPALPQ and/or VLPALP or a functional analogue or derivative thereof is preferred.

The invention also provides use of a signaling molecule for the preparation of a pharmaceutical composition or medicament and methods of treatment for various medical  
15 conditions that are other than use in the preparation of a pharmaceutical composition for the treatment of an immune-mediated-disorder or a method of treatment of an immune-mediated-disorder. For example, the invention provides topical application, for example in an ointment or spray comprising a signal molecule according to the invention, for the prevention or mitigation of skin afflictions, such as eczemas, psoriasis, but also of skin  
20 damage related to over-exposure to UV-light.

Also, use is contemplated in palliative control, whereby a gene related to prostaglandin synthesis is modulated such that COX2 pathways are effected.

Furthermore, the invention also provides use of a signaling molecule for the preparation of a pharmaceutical composition or medicament and methods of treatment for  
25 various medical conditions that are other than use in the preparation of a pharmaceutical composition for the treatment of wasting syndrome, such as the treatment of particular individuals that are suffering from infection with HIV or a method of treatment of wasting syndrome of such individuals.

In one embodiment, the invention provides the use of a signaling molecule  
30 according to the invention for the preparation of a pharmaceutical composition or medicament for modulating angiogenesis or vascularization, in particular during embryonal development, or after transplantation to stimulate vascularization into the

transplanted organ or inhibit it in a later phase. Signaling molecules that effect angiogenesis are disclosed herein in the detailed description.

Use as provided herein also comprises regulating TNF-alpha receptor (e.g., CD27) expression on cells, thereby modulating the relative ratios and/or cytokine activity of lymphocyte, dendritic or antigen presenting cell subset-populations in a treated individual. As for example described in the detailed description, particular oligopeptide according to the invention is capable of down-regulating CD27 expression on cells of the T-cell lineage.

Down-regulating TNF-alpha itself is also particularly useful in septic-shock-like conditions that not only display increased TNF-alpha activity but display further release of other inflammatory compounds, such as NO. NO production is a central mediator of the vascular and inflammatory response. Our results show that inflammatory cells like macrophages stimulated with an inflammatory active compound such as LPS produce large amounts of NO. However, these cells co-stimulated with most of the NMPF peptides (NMPF peptide 1 to 14, 43 to 66 and 69), even in a very low dose (1 pg/ml), inhibited production of NO. Typical septic-shock-like conditions that can preferably be treated by down-regulating TNF-alpha and NO production comprise disease conditions such as those caused by *Bacillus anthracis* and *Yersinia pestis* toxins or infections with these micro-organisms likely involved in bio-terrorism. Anthrax toxin is produced by *Bacillus anthracis*, the causative agent of anthrax, and is responsible for the major symptoms of the disease. Clinical anthrax is rare, but there is growing concern over the potential use of *B. anthracis* in biological warfare and terrorism. Although a vaccine against anthrax exists, various factors make mass vaccination impractical. The bacteria can be eradicated from the host by treatment with antibiotics, but because of the continuing action of the toxin, such therapy is of little value once symptoms have become evident. Thus, a specific inhibitor of the toxin's action will prove a valuable adjunct to antibiotic therapy. The toxin consists of a single receptor-binding moiety, termed protective antigen (PA), and two enzymatic moieties, termed edema factor (EF) and lethal factor (LF). After release from the bacteria as nontoxic monomers, these three proteins diffuse to the surface of mammalian cells and assemble into toxic, cell-bound complexes.

Cleavage of PA into two fragments by a cell-surface protease enables the fragment that remains bound to the cell, PA<sub>63</sub>, to heptamerize and bind EF and LF with high

affinity. After internalization by receptor-mediated endocytosis, the complexes are trafficked to the endosome. There, at low pH, the PA moiety inserts into the membrane and mediates translocation of EF and LF to the cytosol. EF is an adenylate cyclase that has an inhibitory effect on professional phagocytes, and LF is a protease that acts specifically on  
5 macrophages, causing their death and the death of the host.

Down-regulating TNF-alpha itself, and/or a receptor for TNF-alpha, as is herein also provided, is also beneficial in individuals with Chagas cardiomyopathy.

Also, use of a signaling molecule according to the invention for the preparation of a pharmaceutical composition for modulation of vascularization or angiogenesis in  
10 wound repair, in particular of burns, is herein provided. Also, use of a pharmaceutical composition as provided herein is provided in cases of post-operative physiological stress, whereby not only vascularization will benefit from treatment, but the general well-being of the patient is improved as well.

Another use of a signaling molecule according to the invention comprises its  
15 use for the preparation of another pharmaceutical composition for the treatment of cancer. Such a pharmaceutical composition preferably acts via modulating and up-regulating apoptotic responses that are classically down-regulated by NF-kappaB activity. Inhibiting the activity with a signaling molecule according to the invention allows for increased cell death of tumorous cells. Another anti-cancerous activity of a signaling molecule as  
20 provided herein comprises down-regulation of c-myb, in particular in the case of hematopoietic tumors in humans. In this context, down-regulation of 14.3.3 protein is also provided.

A further use of a signaling molecule according to the invention comprises its use for the preparation of a further pharmaceutical composition for the treatment of cancer.  
25 Such a pharmaceutical composition preferably acts via modulating and down-regulating transferrin receptor availability, in particular on tumorous cells. Transferrin receptors are classically up-regulated by NF-kappaB activity. Inhibiting the activity with a signaling molecule according to the invention allows for reduced iron up-take and increased cell death of tumorous cells. In particular erythroid and thromboid cells are susceptible to the  
30 treatment.

Yet a further use of a signaling molecule according to the invention comprises its use for the preparation of yet another pharmaceutical composition for the treatment of

cancer, in particular of cancers that are caused by viruses, such as is the case with retroviral-induced malignancies and other viral-induced malignancies. Such a pharmaceutical composition preferably acts via modulating and down-regulating cell-proliferative responses that are classically up-regulated by virus-induced transcriptional or  
5 NF-kappaB activity. Inhibiting the activity with a signaling molecule according to the invention allows for decreased proliferation and increased cell death of tumorous cells. Such a pharmaceutical composition may also act via modulating angiogenic responses induced by IL-8, whereby for example inhibition of IL-8 expression via inhibition of transcription factor AP-1 or NF-kappaB expression results in the inhibition of  
10 vascularization dependent tumor growth.

Furthermore, the invention provides the use of a signaling molecule for the preparation of a pharmaceutical composition for optimizing human or animal fertility and embryo survival, and a method for optimizing fertility and embryo survival. In particular, the invention provided for a method and composition allowing the down-regulation of  
15 TNF-alpha in the fertilized individual, optimally in combination with a composition and method for up-regulating IL-10 in the individual. Such a composition and method finds immediate use in both human and veterinary medicine.

Also, the invention provides the use of a signaling molecule for the preparation of a pharmaceutical composition for modulating the body weight of an individual, in particular by modulating gene expression of a gene under influence of peroxisome proliferator-activated receptor gamma (PPARgamma) activation and lipid metabolism by applying a signaling molecule according to the invention, and a method for modulating bodyweight comprising providing an individual with a signaling molecule according to the invention.

25 A further use of a signaling molecule as provided herein lies in the modulation of expression of a gene in a cultured cell. Such a method as provided herein comprises subjecting a signaling molecule according to the invention to the cultured cell. Proliferation and/or differentiation of cultured cells (cells having been or being under conditions of in vitro cell culture known in the art) can be modulated by subjecting the  
30 cultured cell to a signaling molecule according to the invention. It is contemplated that for example research into proliferation or differentiation of cells, such as stem-cell research will benefit greatly from understanding that a third major way of effecting gene

modulation exists, and considering the ease of application of synthetic peptides , and analogues or derivatives thereof.

Furthermore, it is contemplated that a signaling molecule as provided herein finds an advantageous use as a co-stimulatory substance in a vaccine, accompanying  
5 modern day adjuvants or replacing the classically used mycobacterial adjuvants, especially considering that certain mycobacteria express hCG-like proteins, of which it is now postulated that these bacteria have already made use of the this third pathway found in gene modulation as provided herein by providing the host with breakdown products mimicking the signaling molecules identified herein. Treatment and use of the  
10 compositions as provided herein is not restricted to animals only, plants and other organisms are also subject to this third pathway as provided herein. Furthermore, now that the existence of such a pathway has been demonstrated, it is herein provided to make it subject of diagnosis as well, for example to determine the gene modulatory state of a cell in a method comprising determining the presence or absence of a signaling molecule as  
15 provided herein or determining the presence or absence of a protease capable of generating such a signaling molecule from a (preferable endogenous) protein.

#### BRIEF DESCRIPTION OF FIGURES

Figures 1-2. Bone marrow (BM) cell yield of treated BALB/c mice (n=6). BM  
20 cells were isolated from treated mice and cultured *in vitro* in the presence of rmGM-CSF for nine days. These figures show cell yield after nine days of culture of BM cells isolated from mice treated with PBS, LPS or LPS in combination with NMPF peptides 4, 46, 7 and 60. In these figures, cell yield is expressed in relative percentage of cells compared to PBS. Each condition consists of 6 Petri dishes and results shown in these figures are  
25 representative of 6 dishes. Differences of  $\geq 20\%$  were considered significant and line bars represent significant data as compared to LPS control group. A representative experiment is shown. Findings involving all experimental conditions were entirely reproduced in 3 additional experiments.

Figure 3. Effect of *in vivo* treatment on MHC-II expression on CD11c<sup>+</sup> cells. Bone  
30 marrow (BM) cells were isolated from treated BALB/c mice (n=6) and cultured *in vitro* in the presence of rmGM-CSF for nine days. This figure shows MHC-II expression expressed in mean fluorescence intensity (MFI) after nine days of culturing of BM cells isolated from

PBS, LPS or LPS in combination with NMPF. Each condition consists of 6 Petri dishes and results shown in these figures are representative of 6 dishes. Differences of  $\geq 20\%$  were considered significant and line bars represent significant data as compared to LPS control group. A representative experiment is shown. Findings involving all experimental conditions were entirely reproduced in 3 additional experiments.

Figures 4-7. Bone marrow (BM) cell yield of *in vitro* treated BM cultures. BM cells from BALB/c mice (n=3) were cultured *in vitro* and treated with either PBS, LPS (t=6), NMPF 4, 7, 46, 60 (t=0 or t=6) or combination of NMPF with LPS (t=6), in the presence of rmGM-CSF for nine days. These figures show cell yield expressed in relative percentage of cells compared to PBS after nine days of culture of BM cells. Each condition consists of 6 Petri dishes and results shown in these figures are representative of 6 dishes. Differences of  $\geq 20\%$  were considered significant. Line bars represent significant data as compared to LPS control group and dotted bars represent significant data as compared to PBS group. A representative experiment is shown. Findings involving all experimental conditions were entirely reproduced in 3 additional experiments.

Figure 8-11. Effect of *in vitro* treatment on MHC-II expression on CD11c<sup>+</sup> cells. BM cells from BALB/c mice (n=3) were cultured *in vitro* and treated with either PBS, LPS (t=6), NMPF 4, 7, 46, 60 (t=0 or t=6 days) or combination of NMPF with LPS (t=6 days), in the presence of rmGM-CSF for nine days. These figures show MHC-II expression expressed in mean fluorescence intensity (MFI) of CD11c positive cells after nine days of culturing of BM cells. Each condition consists of 6 Petri dishes, and results shown in these figures are representative of 6 dishes. Differences of  $\geq 20\%$  were considered significant. Line bars represent significant data as compared to LPS control group and dotted bars represent significant data as compared to PBS group. A representative experiment is shown. Findings involving all experimental conditions were entirely reproduced in 3 additional experiments.

Figure 12-15. Bone marrow (BM) cell yield of treated BALB/c mice (n=6). BM cells were isolated from treated mice and cultured *in vitro* in the presence of rmGM-CSF for nine days. These figures show cell yield after nine days of culture of BM cells in suspension (unattached) and attached to Petri dish (attached). BM cells were isolated from mice treated with PBS, LPS or LPS in combination with different NMPF peptides. In these figures cell yield is expressed in relative percentage of cells compared to PBS. Each

condition consists of 6 Petri dishes and results shown here are representative of 6 dishes. Differences of  $\geq 20\%$  were considered significant and line bars represent significant data as compared to LPS control group. A representative experiment is shown. Findings involving all experimental conditions were entirely reproduced in 3 additional experiments.

Figures 16-17. Bone marrow (BM) cell yield of *in vitro* treated BM cultures from NOD mice. BM cells from 15 weeks old female NOD mice ( $n=3$ ) were cultured *in vitro* and treated with either PBS or NMPF in the presence of rmGM-CSF for nine days. These figures show cell yield after nine days of culture of BM cells in suspension (unattached) and attached to Petri dishes (attached). In these figures cell yield is expressed in relative percentage of cells compared to PBS. Each condition consists of 6 Petri dishes and results shown here are representative of 6 dishes. Differences of  $\geq 20\%$  were considered significant and dotted bars represent significant data as compared to PBS control group. A representative experiment is shown. Findings involving all experimental conditions were entirely reproduced in 3 additional experiments.

Figures 20-32. In vivo treatment of fertilised chicken eggs with NMPF and the effect of NMPF on angiogenesis. Fertile chicken eggs (day 0) were treated with either PBS, NMPF, VEGF or VEGF in combination with NMPF. Ten eggs were injected for every condition. On day 8 of incubation, the embryos were removed from the eggs and were placed in a 100-mm Petri dish. The embryo and the blood vessels were photographed in vivo with the use of a microscope. Of each egg one overview picture was taken and 4 detail pictures of the blood vessels were taken. Quantification of angiogenesis (vessels branches) was accomplished by counting the number of blood vessels branches. Quantification of this vasculogenesis was accomplished by measuring the blood vessel thickness. The number of blood vessel branches and vessel thickness was measured in the pictures and were correlated to a raster (in the pictures) of  $10\text{mm}^2$  for comparison. The mean number of branches and the mean blood vessel thickness of each condition ( $N=10$ ) were calculated and compared to either the PBS or VEGF controls using a Student's T-test. Line bars represent significant ( $p<0.05$ ) data as compared to PBS control group and dotted bars represent significant ( $p<0.05$ ) data as compared to VEGF group. Figures 20-30 show the effect of NMPF on vessel branches. Figures 31-32 show the effect of NMPF on vessel thickness.

Figure 33. Detection of NF- $\kappa$ B via EMSA. This figure shows the presence of NF- $\kappa$ B in the nuclear extracts of RAW264.7 cells treated with LPS or NMPF in combination with LPS for 4 hours. Numbers 1-13 correspond to nuclear extracts from cells treated with NMPF and LPS. CTL corresponds to nuclear extracts from cells treated with LPS only. Specificity of the radioactively labeled NF- $\kappa$ B probe is shown by competition with the unlabeled oligonucleotide (u1,u2,u3) in three different concentrations (1x, 10x, 100x) with nuclear extracts of CTL and *olg* corresponds to samples containing only labeled oligonucleotide (without nuclear extract). Description: (NMPF-1)VLPALPQVVC, (NMPF-2)LQGVLPALPQ, (NMPF-3)LQG, (NMPF-4)LQGV, (NMPF-5)GVLPALPQ, (NMPF-6)VLPALP, (NMPF-7)VLPALPQ, (NMPF-8)GVLPALP, (NMPF-9)VVC, (NMPF-11)MTRV, (NMPF-12)MTR

Figure 34. HPLC chromatograph (wave length 206) in which data profile obtained from the nuclear protein extracts of LPS and LPS in combination with NMPF stimulated RAW264.7 cells are overlaid.

Figure 35. MSn analysis of NMPF-4 peptide.

Figure 36. MSn analysis of fraction from nuclear extract of LPS stimulated RAW264.7 cells. Upper panel shows full spectrum of the fraction and lower panel shows the MS/MS spectrum of mass 413.13.

Figure 37. MSn analysis of fraction from nuclear extract of LPS in combination with NMPF-4 stimulated RAW264.7 cells. Upper panel shows full spectrum of the fraction and lower panel shows the MS/MS spectrum of mass 416.07.

Figures 38 – 49 Effect of NMPF on septic shock syndrome in Rhesus monkeys. On the time point 70 minutes *E. coli* was infused and at the end of *E. coli* infusion (time point 190 minutes) the antibiotic Baytril was injected. The control monkey (monkey 429) was treated with 0.9% NaCl at the time point of 100 minutes, whereas the NMPF treated monkeys (monkey 459 and 427) received the NMPF treatment at the same time point as the control monkey. Heart rate (beats per minute), blood pressure (mmHg), difference between systolic and diastolic blood pressure and blood oxygen concentration (saturation in %) of the control monkey 429 (figures 38-41), NMPF treated monkey 459 (figures 42-45) and 427 (figures 46-49) in the time (minutes) during the experiment are shown.



Figure 50 These figures show the NO production of LPS (10 µg/ml) stimulated RAW 264.7 macrophages co-stimulated with different NMPF peptides (1 pg/ml).

Figure 51 The figure shows the NO production of LPS (10 µg/ml) stimulated RAW 264.7 macrophages co-stimulated with different NMPF peptides with three different  
5 concentrations.

Figure 52 This figure shows the percentage of diabetic NOD mice treated for 2 weeks with the various NMPF peptides

Figure 53 These figures show the performed glucose tolerance test (GTT) in NOD mice treated with NMPF peptides (A), and fasting blood glucose levels (B).

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#### DETAILED DESCRIPTION OF THE INVENTION

Cells react to environmental and intrinsic changes, which they perceive through extracellular and inter- as well as intracellular signals. The nature of these signals can be either for example physical or chemical. Moreover, different classes of molecules present  
15 in blood react to each other and induce cascade of reaction that have direct effect on other molecules and/or eventually lead to cellular responses, for example complement system and blood coagulation proteins.

Many genes are regulated not by a signaling molecule that enters the cells but by molecules that bind to specific receptors on the surface of cells for example receptors with  
20 enzymatic activity (receptor tyrosine kinases, receptor-like protein tyrosine phosphatases, receptor serine/threonine kinases, histidine kinases, guanylyl cyclases) and receptors without enzymatic activity (cytokine receptors, integrins, G-protein-coupled receptors). Interaction between cell-surface receptors and their ligands can be followed by a cascade of intracellular events that modulate one or more intracellular transducing proteins,  
25 including variations in the intracellular levels of so-called second messengers (diacylglycerol,  $\text{Ca}^{2+}$ , cyclic nucleotides, inositol(1,4,5) trisphosphate, phosphatidylinositol(3,4,5) trisphosphate, phosphatidylinositol transfer protein (PITP)). This leads to the activation or inhibition of a so-called 'effector protein'. The second messengers in turn lead to changes in protein for example protein phosphorylation through  
30 the action of cyclic AMP, cyclic GMP, calcium-activated protein kinases, or protein kinases (for example AGC group serine/threonine protein kinases, CAMK group serine/threonine protein kinases, CMGC group serine/threonine kinases, protein tyrosine

kinase group, or others like MEK/Ste7p). Phosphorylation by protein kinases is one of the regulatory mechanisms in signal transmission that modulate different cellular pathways such as Ras/MAPK pathway, MAP kinase pathway, JAK-STAT pathway, wnt-pathway. In many instances this all results in altered gene expression (for example genes for the regulation of other genes, cell survival, growth, differentiation, maturation, functional activity).

Many of the responses to binding of ligands to cell-surface receptors are cytoplasmatic and do not involve immediate gene activation in the nucleus. In some instances a pre-existing inactive transcription factor following a cell-surface interaction is activated that lead to immediate gene activation . For example the protein NF-kappaB, which can be activated within minutes by a variety of stimuli, including membrane receptors (for example pattern recognition receptors like Toll-like receptor binding to pathogen-associated molecular patterns), inflammatory cytokines such as TNF- $\alpha$ , IL-1, T-cell activation signals, growth factors and stress inducers.

Our genomic experiment with NMPF peptide LQGV showed very immediate effects on signal transduction and gene regulation since the cells were treated with the peptide for only four hours. In this short period of time LQGV down-regulated at least 120 genes and up-regulated at least 6 genes in the presence of a strong stimulator (PHA/IL-2 stimulated T-cell line (PM1)), demonstrating the profound effect on signaling molecule according to the invention and modulatory effect on gene expression. The genes affected by LQGV include onco-genes, genes for transcription factors, intracellular enzymes, membrane receptors, intracellular receptors, signal transducing proteins (for example kinases) and some genes for unknown molecules. This shows that LQGV as an example of the synthetic signaling molecule (oligopeptide or functional analogue or derivative thereof) as described here, has broad spectrum of effects at different extracellular and intracellular levels. In addition, our HPLC/MS data have shown the presence of LQGV in the nucleus of a macrophage cell line (RAW267.4) within half hour and also indicates the direct effects on DNA level as well as at intracellular level which is further supported by NF-kappaB experiments. The ultimate modulatory effect of LQGV is dependent on, for example, type of the cell, differentiation and maturation status of the cell, the functional status and the presence of other regulatory molecules. This was evident by shock experiment in which different NMPF peptides had similar or different effects on the disease. Same results were

obtained with DC, fertilized chicken egg experiments, and CAO experiments; NMPF effects were dependent on type of co-stimulation (GM-CSF alone or in combination with LPS, or VEGF) and time of the treatment. Due to this NMPF have the ability to modulate cellular responses at different levels.

5

## EXAMPLES

### MATERIAL AND METHODS

#### Peptide synthesis

The peptides as mentioned in this document such as LQG, AQG, LQGV, AQGV, LQGA, VLPALP, ALPALP, VAPALP, ALPALPQ, VLPAAPQ, VLPALAQ, LAGV, 10 VLAALP, VLPALA, VLPALPQ, VLAALPQ, VLPALPA, GVLPALP, VVCNYRDVRFESIRLPGCPRGVNPVVSYAVALSCQCAL, RPRCRPINATLAVEKEGCPVCITVNTTICAGYCPT, SKAPPPSLPSPSRLPGPS, LQGVLPALPQVVC, SIRLPGCPRGVNPVVS, LPGCPRGVNPVVS, LPGC, MTRV, MTR, VVC were prepared by solid-phase synthesis (Merrifield, 1963) using the 15 fluorenylmethoxycarbonyl (Fmoc)/tert-butyl-based methodology (Atherton, 1985) with 2-chlorotrityl chloride resin (Barlos, 1991) as the solid support. The side-chain of glutamine was protected with a trityl function. The peptides were synthesized manually. Each coupling consisted of the following steps: (i) removal of the alpha-amino Fmoc-protection by piperidine in dimethylformamide (DMF) (ii) coupling of the Fmoc amino acid (3 eq) 20 with diisopropylcarbodiimide (DIC)/1-hydroxybenzotriazole (HOBt) in DMF/N-methylformamide (NMP) (iii) capping of the remaining amino functions with acetic anhydride/diisopropylethylamine (DIEA) in DMF/NMP. Upon completion of the synthesis, the peptide resin was treated with a mixture of trifluoroacetic acid (TFA)/H<sub>2</sub>O/triisopropylsilane (TIS) 95:2.5:2.5. After 30 minutes TIS was added until 25 decoloration. The solution was evaporated in vacuo and the peptide precipitated with diethylether. The crude peptides were dissolved in water (50-100 mg/ml) and purified by reverse-phase high-performance liquid chromatography (RP-HPLC). HPLC conditions were: column: Vydac TP21810C18 (10 x 250 mm); elution system: gradient system of 0.1% TFA in water v/v (A) and 0.1% TFA in acetonitrile (ACN) v/v (B); flow rate 6 30 ml/min; absorbance was detected from 190-370 nm. There were different gradient systems used. For example for peptides LQG and LQGV: 10 minutes 100% A followed by linear gradient 0-10% B in 50 minutes. For example for peptides VLPALP and VLPALPQ: 5

minutes 5% B followed by linear gradient 1% B/minute. The collected fractions were concentrated to about 5 ml by rotation film evaporation under reduced pressure at 40°C. The remaining TFA was exchanged against acetate by eluting two times over a column with anion exchange resin (Merck II) in acetate form. The elute was concentrated and lyophilised in 28 hours. Peptides later were prepared for use by dissolving them in PBS.

### Transcription factor experiment

*Macrophage cell line.* The RAW 264.7 macrophages, obtained from American Type Culture Collection (Manassas, VA), were cultured at 37°C in 5% CO<sub>2</sub> using DMEM containing 10% FBS and antibiotics (100 U/ml of penicillin, and 100 µg/ml streptomycin). Cells (1 x10<sup>6</sup>/ml) were incubated with peptide (10 µg/ml) in a volume of 2 ml. After 8 h of cultures cells were washed and prepared for nuclear extracts.

*Nuclear extracts.* Nuclear extracts and EMSA were prepared according to Schreiber et al. Methods (Schriber et al. 1989, Nucleic Acids Research 17). Briefly, nuclear extracts from peptide stimulated or nonstimulated macrophages were prepared by cell lysis followed by nuclear lysis. Cells were then suspended in 400 µl of buffer (10 mM HEPES (pH 7.9), 10 mM KCl, 0.1 mM KCL, 0.1 mM EDTA, 0.1 mM EGTA, 1 mM DTT, 0.5 mM PMSF and protease inhibitors), vigorously vortexed for 15 s, left standing at 4°C for 15 min, and centrifuged at 15,000 rpm for 2 min. The pelleted nuclei were resuspended in buffer (20 mM HEPES (pH 7.9), 10% glycerol, 400 mM NaCl, 1 mM EDTA, 1mM EGTA, 1 mM DTT, 0.5 mM PMSF and protease inhibitors) for 30 min on ice, then the lysates were centrifuged at 15,000 rpm for 2 min. The supernatants containing the solubilized nuclear proteins were stored at -70°C until used for the Electrophoretic Mobility Shift Assays (EMSA).

*EMSA.* Electrophoretic mobility shift assays were performed by incubating nuclear extracts prepared from control (RAW 264.7) and peptide treated RAW 264.7 cells with a 32P-labeled double-stranded probe (5' AGCTCAGAGGGGGACTTTCCGAGAG 3') synthesized to represent the NF-kappaB binding sequence. Shortly, the probe was end-labeled with T4 polynucleotide kinase according to manufacturer's instructions (Promega, Madison, WI). The annealed probe was incubated with nuclear extract as follows: in EMSA, binding reaction mixtures (20 µl) contained 0.25 µg of poly(dI-dC) (Amersham Pharmacia Biotech) and 20,000 rpm of 32P-labeled DNA probe in binding buffer

consisting of 5 mM EDTA, 20% Ficoll, 5 mM DTT, 300 mM KCl and 50 mM HEPES. The binding reaction was started by the addition of cell extracts (10 µg) and was continued for 30 min at room temperature. The DNA-protein complex was resolved from free oligonucleotide by electrophoresis in a 6% polyacrylamide gel. The gels were dried and  
5 exposed to x-ray films.

### Apo E experiments

Apolipoprotein E (apo E) deficiency is associated with a series of pathological conditions including dyslipidemia, atherosclerosis, Alzheimer's disease, increase body  
10 weight and shorter life span. Inheritance of different alleles of the POLYMORPHIC *apoE* gene is responsible for 10% of the variation in plasma cholesterol in most populations. Individuals HOMOZYGOUS for one variant. *apoE2*, can develop type III  
dysbetalipoproteinaemia if an additional genetic or environmental factor is present. Some  
15 much rarer alleles of *apoE* produce dominant expression of this disorder in heterozygous individuals. ApoE, is a ligand for the LDL receptor and its effects on plasma cholesterol are mediated by differences in the affinity of the LDL receptor for lipoproteins carrying variant apoE proteins. The factors that regulate *apoE* gene transcription have been investigated extensively by the expression of gene constructs in transgenic mice and involve complex interactions between factors that bind elements in the 5' promoter region,  
20 in the first intron and in 3' regions many kilobases distant from the structural gene. Deletion of the apo E gene is associated with changes in lipoprotein metabolism [plasma total cholesterol), HDL cholesterol, HDL/TC, and HDL/LDL ratios, esterification rate in apo B-depleted plasma, plasma triglyceride, hepatic HMG-CoA reductase activity, hepatic cholesterol content, decreased plasma homocyst(e)ine and glucose levels, and severe  
25 atherosclerosis and cutaneous xanthomatosis.

## RESULTS

### NF-kB experiments

The transcription factor NF-kB participates in the transcriptional regulation of a  
30 variety of genes. Nuclear protein extracts were prepared from LPS and peptide treated RAW264.7 cells or from LPS treated RAW264.7 cells. In order to determine whether the peptide modulates the translocation of NF-kB into the nucleus, on these extracts EMSA

was performed. Figure 33 shows the amount of NF-kB present in the nuclear extracts of RAW264.7 cells treated with LPS or LPS in combination with peptide for 4 hours. Here we see that indeed some peptides are able to modulate the translocation of NF-kB since the amount of labeled oligonucleotide for NF-kB is reduced. In this experiment peptides that show the modulation of translocation of NF-kB are: (NMPF-1)VLPALPQVVC, (NMPF-2)LQGVLPALPQ, (NMPF-3)LQG, (NMPF-4)LQGV, (NMPF-5)GVLPALPQ, (NMPF-6)VLPALP, (NMPF-7)VLPALPQ, (NMPF-8)GVLPALP, (NMPF-9)VVC, (NMPF-11)MTRV, (NMPF-12)MTR.

## 10 Nuclear location of peptide experiment

Reverse-phase high-performance liquid chromatography (RP-HPLC) method was used to prove the presence of synthetic oligo-peptide in the nuclear extracts. We used a Shimadzu HPLC system equipped with Vydac monomeric C18 column (column 218MS54, LC/MS C18 reversed phase, 300A, 5  $\mu$ m, 4.6mm ID x 250mm L); elution system: gradient system of 0.01% TFA and 5% acetonitrile (CAN) in water v/v (A) and 0.01% TFA in 80% acetonitrile (ACN) v/v (B); flow rate 0.5 ml/min; absorbance was detected from 190-370 nm. The gradient time program was as follows:

Time (min)	Buffer B concentration
0.01	0
5.0	0
30.0	80
40.0	100
60.0	100
65.0	0
70.0	0

The elution time of peptide LQGV was determined by injecting 2  $\mu$ g of the peptide in a separate run. Mass spectrometry (MS) analysis of fraction which contained possible NMPF-4 (LQGV) (elution time was determined by injecting the peptide in the same or separate run) was performed on LCQ Deca XP (Thermo Finnigan).

## RESULTS

### Nuclear location of peptide experiment

The nuclear protein extracts used in EMSA experiments were also checked for the presence of LQGV by means of HPLC and MS. Figure 34 shows HPLC chromatograph (wave length 206) in which data profile obtained from the nuclear protein extracts of LPS and LPS in combination with NMPF-4 (LQGV) stimulated RAW264.7 cells are overlaid.

5 This figure also show the presence or absence of number of molecule signals in the nuclear extracts of oligopeptide+LPS treated cells as compared to nuclear extracts of LPS treated cells. Since HPLC profile of LQGV showed that the peptide elutes at around 12 minutes (data not shown), fraction corresponding to region 10-15 minutes was collected and analysed for the presence of this peptide in MS.

10 The molecular weight of this peptide is around 416 dalton. Besides 416 mass figure 35 shows some other molecular weights. This is to be explained by the high concentration of the peptide which induces the formation of dimers and sodium-adducts ( $m/z$  416- $[M+H]^+$ , 438- $[M+Na]^+$ , 831- $[2M+H]^+$ , 853- $[2M+Na]^+$ , 1245- $[3M+H]^+$ , 1257- $[3M+Na]^+$ ). Figure 36 shows the MS results of 10-15 min. fraction of nuclear extract  
15 obtained from LPS stimulated cells. These results show the absence of 416 dalton mass, while figure 37 shows the presence of 416 dalton mass of which the MSn data (figure 37) and MS-sequence confirm the presence of LQGV peptide in the nuclear protein extract obtained from LQGV+LPS stimulated RAW264.7 cells.

## 20 **Endotoxin shock model (Sepsis)**

*Sepsis.* For the endotoxin model, BALB/c mice were injected i.p. with 8-9 mg/kg LPS (*E. coli* 026:B6; Difco Lab., Detroit, MI, USA). Control groups (PBS) were treated with PBS i.p. only. To test the effect of NMPF from different sources (synthetic, commercial hCG preparation [c-hCG]), we treated BALB/c with a dose of 300-700 IU of  
25 different hCG preparations (PG23;Pregnyl batch no. 235863, PG25; Pregnyl batch no. 255957) and with synthetic peptides (5 mg/kg) after two hours of LPS injection. In other experiments BALB/c mice were injected i.p. either with 10 mg/kg or with 11 mg/kg LPS (*E. coli* 026:B6; Difco Lab., Detroit, MI, USA). Subsequently mice were treated after 2 hours and 24 hours of LPS treatment with NMPF peptides.

30 *Semi-quantitative sickness measurements.* Mice were scored for sickness severity using the following measurement scheme:

- 1 Percolated fur, but no detectable behaviour differences compared to normal mice.
- 2 Percolated fur, huddle reflex, responds to stimuli (such as tap on cage), just as active during handling as healthy mouse.
- 5 3 Slower response to tap on cage, passive or docile when handled, but still curious when alone in a new setting.
- 4 Lack of curiosity, little or no response to stimuli, quite immobile.
- 5 Laboured breathing, inability or slow to self-right after being rolled onto back (moribund)
- 10 6 Sacrificed

## RESULTS

### Endotoxin shock model (Sepsis)

*Sepsis experiments.* To determine the effect of synthetic peptides (NMPF) in high-dose LPS shock model, BALB/c mice were injected intraperitoneally with different doses of LPS and survival was assessed daily for 5 days. In this experiment (for the LPS endotoxin model) BALB/c mice were injected i.p. with 8-9 mg/kg LPS (E. coli 026:B6; Difco Lab., Detroit, MI, USA). Control groups (PBS) were treated with PBS i.p. only. We treated BALB/c mice with a dose of 300-700 IU of different hCG preparations (PG23; Pregnyl batch no. 235863, PG25; Pregnyl batch no. 255957) or with peptides (5 mg/kg) after two hours of LPS injection.

These experiments showed (Table 1) that NMPF peptides 4, 6, 66 and PG23 inhibited shock completely (all mice had in first 24 hours sickness scores not higher than 2; shortly thereafter they recovered completely and had sickness scores of 0), while peptides 2, 3 and 7 accelerated shock (all mice had in first 24 hours sickness scores of 5 and most of them died, while the control mice treated with LPS+PBS had sickness scores of 3-4 in first 24 hours and they most of them died after 48 hours with sickness scores of 5 (17% survival rate at 72 hours). In addition, peptides 1, 5, 8, 9, 11, 12, 13, 14 and 64 showed in number of different experiments variability in effectiveness as well as in the kind (inhibitory vs accelerating) of activity. This variability is likely attributable to the rate of breakdown of the various peptides, and the different effects the various peptides and their breakdown products have *in vivo*. In addition, these experiments also showed the variability in anti-



shock activity in c-hCG preparations that is likely attributable to the variation in the presence of anti-shock and shock accelerating NMPF. Visible signs of sickness were apparent in all of the experimental animals, but the kinetics and obviously the severity of this sickness were significantly different. These data are representative of at least 10  
5 separate experiments.

In Table 2 we see the effect of ALA-replacement (PEPSCAN) in peptide LQG, LQGV, VLPALP, VLPALPQ in septic shock experiments. We conclude, that the change in even one amino acid by a neutral amino acid can lead to different activity. So, genomic differences as well as polymorphism in these peptides can regulate the immune response  
10 very precise. Derivatives of these peptides, for example (but not limited to) by addition of classical and non-classical amino acids or derivatives that are differentially modified during or after synthesis, for example benzylation, amidation, glycosylation, proteolytic cleavage, linkage to an antibody molecule or other cellular ligand etc. could also lead to a better effectiveness of the activity.

15 To determine whether treatment of BALB/c mice with NMPF inhibit septic shock at different stages of disease, synthetic peptides (NMPF) were injected i.p. at 2 and 24 hours after the induction of septic shock with high dose LPS (10 mg/kg).

As shown in Tables 3 and 4., control mice treated PBS after the shock induction, reached a sickness score of 5 at 14 and 24 hours, and remained so after the second injection  
20 with PBS. The survival rate in control group mice was 0% at 48 hours. In contrast to control mice, mice treated with NMPF 9, 11, 12, 43, 46, 50 and 60 reached a maximum sickness score of 2-3 at 24 hours after the induction of septic shock and further reached a maximum sickness score of 1-2 at 48 hours after the second injection of NMPF. In addition, mice treated with NMPF 5, 7, 8, 45, 53 and 58 reached a sickness score of 5 and  
25 after the second injection with NMPF all mice returned to a sickness score of 1-2 and survival rates in NMPF groups were 100%. Mice treated with NMPF 3 reached sickness scores of 3-4 and the second NMPF injection did save these mice. These experiments show that NMPF peptides have anti-shock activity at different stages of the disease and NMPF have anti-shock activity even at disease stage when otherwise irreversible damage had  
30 been done. This indicates that NMPF have effects on different cellular levels and also have repairing and regenerating capacity.

### Dendritic cells experiments

*Mice.* The mouse strain used in this study was BALB/c (Harlan, Bicester, Oxon, GB). All mice used in experiments were females between 8 and 12 weeks of age. Mice were housed in a specific-pathogen-free facility. The Animal Use Committee at the  
5 Erasmus University Rotterdam, The Netherlands approved all studies.

*In vivo treatment.* At least six mice per group were injected intraperitoneally (i.p) with LPS (10 mg/kg; Sigma). After 2 and 24 hrs of LPS induction, mice were injected i.p. with either NMPF (5 mg/kg) or Phosphate Buffered Saline (PBS), in a volume of 100 µl. LPS induced shock in this model had more than 90% mortality at 48 hrs.

10 *Bone marrow cell culture.* From treated mice, bone-marrow cells were isolated and cultured as follows. BALB/c mice were killed by suffocation with CO<sub>2</sub>. The femurs and tibiae were removed and freed of muscles and tendons under aseptic conditions. The bones were placed in R10 medium (RPMI 1640, supplemented with 50 U/ml penicillin, 50 µg/ml streptomycin, 0.2 M Na-pyruvate, 2 mM glutamine, 50 µM 2-mercaptoethanol and 10%  
15 fetal calf serum (Bio Whittaker, Europe)).

The bones were then cleaned more thoroughly by using an aseptic tissue and were transferred to an ice cold mortar with 2 ml of R10 medium. The bones were crushed with a mortar to get the cells out of the bones. Cells were filtered through a sterile 100 µM filter (Beckton Dickinson Labware) and collected in a 50 ml tube (FALCON). This procedure  
20 was repeated until bone parts appeared translucent.

The isolated cells were resuspended in 10 ml of R10 and 30 ml of Geys medium was added. The cell suspension was kept on ice for 30 minutes, to lyse the red blood cells. Thereafter, the cells were washed twice in R10 medium. Upon initiation of the culture, the cell concentration was adjusted to  $2 \times 10^5$  cells per ml in R10 medium supplemented with  
25 20 ng/ml recombinant mouse Granulocyte Monocyte-Colony Stimulating Factor (rmGM-CSF; BioSource International, Inc., USA) and seeded in 100 mm non-adherent bacteriological Petri dishes (Falcon). For each condition six Petri dishes were used and for further analysis, cells were pooled and analysed as described ahead. The cultures were placed in a 5% CO<sub>2</sub>-incubator at 37°C. Every three days after culture initiation, 10 ml fresh  
30 R10 medium supplemented with rmGM-CSF at 20 ng/ml was added to each dish.

Nine days after culture initiation, non-adherent cells were collected and counted with a Coulter Counter (Coulter).

Alternatively, BM cells from untreated mice were isolated and cultured as described above and were *in vitro* treated with the following conditions: NMPF 4, NMPF 46, NMPF 7, NMPF 60 (20 µg/ml) were added to the culture either at day 0 or day 6 after culture initiation. Or LPS (1 µg/ml) was added to the culture at day 6 with or without the NMPF.

*Immunofluorescence staining.* Cells ( $2 \times 10^5$ ) were washed with FACS-buffer (PBS with 1% BSA and 0.02% sodium azide), and transferred to a round-bottomed 96-well plate (NUNC). The antibodies used for staining were against MHC-II (I-A/I-E) PE and CD11c/CD18 FITC (PharMingen/Becton Dickinson, Franklin Lakes, NJ, USA).

Cells were resuspended in 200 µl FACS-buffer containing both of the antibodies at a concentration of 2.5 ng/µl per antibody. Cells were then incubated for 30 min at 4°C. Thereafter, cells were washed 3 times and finally resuspended in 200 µl FACS-buffer for flow-cytometric analysis in a FACSCalibur flow cytometer (Becton Dickinson, Heidelberg, Germany). All FACS-data were analyzed with CellQuest software (Becton Dickinson, Heidelberg, DE). *Statistical analysis* All differences greater than 20% are considered to be significant.

## RESULTS

### Dendritic cells experiments

*Cell yield of ex vivo bone-marrow cell cultures.* To determine the *in vivo* effect of LPS and NMPF treatment on the cell yield obtained from a nine-day culture of bone-marrow with rmGM-CSF, cells were isolated from the BM of treated mice and cultured, harvested and counted as described. As shown in Figure 1 and 2, the cell yield of the bone-marrow cultures of LPS (10 mg/kg) treated mice is significantly decreased compared to PBS treated mice. Mice treated with NMPF 4, NMPF 7, NMPF 46 and NMPF 60 after LPS shock induction, had a significantly increased cell yield compared to LPS in the presence of rmGM-CSF. In addition, BM cultures from NMPF 46 treated mice gave a significantly increased cell yield even compared to the PBS group.

*Immunofluorescence staining of in vivo treated bone-marrow derived DC.* Culture of BM cells in the presence of rmGM-CSF gave rise to an increased population of cells that are positive for CD11c and MHC-II. Cells positive for these cell membrane markers are bone-marrow derived dendritic cells (DC). DC are potent antigen presenting cells

(APC) and modulate immune responses. In order to determine the maturation state of myeloid derived DC, cells were stained with CD11c and MHC-II.

As shown in Figure 3, the expression of the MHC-II molecule was significantly decreased on CD11c-positive cells from LPS treated mice as compared to the PBS group.

5 This decrease in MHC-II expression was further potentiated by the *in vivo* treatment with NMPF 4 and NMPF 46. However, treatment of mice with NMPF 7 and NMPF 60 significantly increased the expression of the MHC-II molecule even as compared to the PBS group.

*Cell yields of in vitro bone-marrow cell cultures.* To determine the effect of LPS and NMPF *in vitro* on the cell yield of a nine-day culture of bone-marrow cells, we  
10 isolated the BM cells from untreated BALB/c mice and cultured in the presence of rmGM-CSF. In addition to rmGM-CSF, cultures were supplemented with NMPF at either day 0 or day 6 with or without the addition of LPS at day 6.

As shown in Figures 4-7, there is a significant decrease in cell yield in LPS treated  
15 BM cells as compared to PBS. BM cells treated with NMPF 4, 7, 46 or 60 at time point t=0 or t=6 without LPS, showed a significant increase in cell yield as compared to the PBS group. However, BM cell cultures treated with NMPF 4 at time point t=6 showed significant decrease in cell yield as compared to the PBS group and this effect is comparable with the effect of LPS (Figure 4). In addition, BM cells treated with NMPF 4,  
20 7, 46 or 60 at time point t=6 in combination with LPS showed a significant increase in cell yield as compared to the LPS group and even in the group of NMPF 7 the cell yield was significantly increased as compared to the PBS group.

*Immunofluorescence staining of in vitro treated bone-marrow derived DC.* To determine the maturation state of DC, CD11c positive cells were stained for MHC-II  
25 antibody. Figure 7-11 show that there is an opposite effect of LPS on MHC-II expression as compared to *in vivo* experiments, namely, MHC-II expression is significantly increased with LPS treatment *in vitro* as compared to PBS. NMPF 4 with LPS further potentiated the effect of LPS, while NMPF 7 with or without LPS (t=6), significantly inhibited the expression of MHC-II as compared to LPS and PBS, respectively. However, cells treated  
30 with NMPF 46 without LPS (t=0) showed significantly increased expression of MHC-II on CD11c positive cells. Furthermore, no significant differences were found in the group

NMPF 60 with or without LPS on MHC-II expression as compared to LPS and PBS treated cells.

To determine the *in vivo* effect of LPS and NMPF treatment on the cell yield obtained from a nine-day culture of bone-marrow with rmGM-CSF, cells were isolated from the BM of treated mice and cultured, harvested and counted as described. The cell yield of 'attached' cells was significant increased with NMPF 4, 7, 9, 11, 43, 46, 47, 50, 53, 58 60 and even in the group of NMPF 7, 46 and 60 the cell yield was significant increased as compared to the PBS group (figure 14-15). In addition, cell yield of 'un-attached' cells was significant increased with NMPF 4, 7, 9, 11, 46, 50, 53, 58 60 and again in the group of NMPF 46 the cell yield was significant increased as compared to the PBS group (Figures 12-13).

To determine the effect of LPS and NMPF *in vitro* on the cell yield of a nine-day culture of bone-marrow cells of female NOD mice, we isolated the BM cells from untreated NOD mice and cultured in the presence of rmGM-CSF. In addition to rmGM-CSF, cultures were supplemented with NMPF. In these experiments the bone-marrow cell yield of 'un-attached' cells was significant increased with NMPF 1,2,3,4,5,6,7,8,9, 12 and 13 as compared to PBS group and no effect was observed with NMPF 11 (Figure 16). The 'attached' bone-marrow cells of these experiments showed different yield than the 'un-attached' cells, namely there was a significant increased in cell yield in cultures treated with NMPF 3 and 13, while cultures treated with NMPF 2 and 6 showed significant decrease in the cell yield as compared to PBS (Figure 17) (additional results are summarized in Table 5).

### **Coronary Artery Occlusion (CAO) experiments**

*CAO induction and treatment.* NMPF have immunoregulatory effects in chronic inflammatory as well as acute inflammatory mice models. Since certain cytokines like TGF-beta1, TNF-alpha, IL-1 and ROS (reactive oxygen species) have been implicated in irreversible myocardial damage produced by prolonged episodes of coronary artery occlusion and reperfusion *in vivo* that leads to ischaemia and myocardial infarct, we tested the cardio-protective properties of peptides in ad libitum fed male Wistar rats (300 g). The experiments were performed in accordance with the Guiding principles in the Care and Use of Animals as approved by the Council of the American Physiological Society and

under the regulations of the Animal Care Committee of the Erasmus University Rotterdam. Shortly, rats (n=3) were stabilised for 30 minutes followed by i.v. 1 ml of peptide treatment (0.5 mg/ml) in 10 minutes. Five minutes after completion of treatment, rats were subjected to a 60-min coronary artery occlusion (CAO). In the last 5 minutes of CAO, rats were again treated over 10 minutes i.v. with 1 ml of peptide (0.5 mg/ml) followed by 120 minutes of reperfusion (IP). Experimental and surgical procedures are described in detail in Cardiovascular Research 37(1998) 76-81. At the end of each experiment, the coronary artery was re-occluded and was perfused with 10 ml Trypan Blue (0.4%, Sigma Chemical Co.) to stain the normally perfused myocardium dark blue and delineate the nonstained area at risk (AR). The heart was then quickly excised and cut into slices of 1 mm from apex to base. From each slice, the right ventricle was removed and the left ventricle was divided into the AR and the remaining left ventricle, using micro-surgical scissors. The AR was then incubated for 10 min in 37°C Nitro-Blue-Tetrazolium (Sigma Chemical Co.; 1 mg per 1 ml Sorensen buffer, pH 7.4), which stains vital tissue purple but leaves infarcted tissue unstained. After the infarcted area (IA) was isolated from the noninfarcted area, the different areas of the LV were dried and weighed separately. Infarct size was expressed as percentage of the AR. Control rats were treated with PBS.

## RESULTS

### 20 Coronary Artery Occlusion (CAO) experiments

Our CAO data showed that 15 rats in control group treated with only PBS had infarcted area of  $70 \pm 2\%$  (average  $\pm$  standard error) after 60-minutes of CAO followed by 2 hours of reperfusion. While rats treated with peptide VLPALP, LQGV, VLPALPQVVC, LQGVLPALPQ, LAGV, LQAV and MTRV showed infarcted area of  $62 \pm 6\%$ ,  $55 \pm 6\%$ ,  $55 \pm 5\%$ ,  $67 \pm 2\%$ ,  $51 \pm 4\%$ ,  $62 \pm 6\%$  and  $68 \pm 2\%$ , respectively. Here, we see that certain peptides (such as VLPALP, LQGV, VLPALPQVVC, LAGV) have a protective effect on the area at risk for infarction. In addition, peptide LQAV showed smaller infarcted area but in some instances the area was haemorrhagic infarcted. In addition NMPF-64 (LPGCPRGVNPVVS) had also protective effect (35%) in CAO experiment. It is important to note that mice treated with certain above mentioned peptides showed less viscosity of blood. Apart from immunological effect, these peptides may have also effect on blood coagulation system directly or indirectly since there is certain homology with blood

coagulation factors ( for additional results of NMPF peptides see table 5.) So, in both models the circulatory system plays an important role in the pathogenesis of the disease.

### Chicken eggs experiments

5        *In vivo treatment of fertilised chicken eggs with NMPF.* Fertile chicken eggs (Drost Loosdrecht BV, the Netherlands) were incubated in a diagonal position in an incubator (Pas Reform BV, the Netherlands) at 37 °C and 32% relative humidity.

10        Solutions of NMPF peptides (1mg/ml) and VEGF were made in PBS. At least ten eggs were injected for every condition. The treatment was performed as follows: on day 0 of incubation, a hole was drilled into the eggshell to open the air cell. A second hole was drilled 10 mm lower and right from the first hole for injection. The holes in the eggshell were disinfected with jodium. The NMPF peptides (100 µg/egg) and/or VEGF (100 ng/ml) were injected in volume of 100 µl. The holes in the eggshell were sealed with tape (Scotch Magic™ Tape, 3M) and the eggs were placed into the incubator.

15        *Quantification of angiogenesis.* On day 7 of incubation, the eggs were viewed under an UV lamp to check if the embryos were developing in a normal way and the dead embryos were counted. On day 8 of incubation, the embryos were removed from the eggs by opening the shell at the bottom of the eggs. The shell membrane was carefully dissected and removed. The embryos were placed in a 100-mm Petri dish. The embryo and the blood vessels were photographed (Nikon E990, Japan) *in vivo* with the use of a microscope (Zeiss Stemi SV6, Germany). One overview picture was taken and 4 detail pictures of the blood vessels were taken. Only eggs with vital embryos were evaluated.

20        *Data analysis.* Quantification of angiogenesis was accomplished by counting the number of blood vessels branches. Quantification of vasculogenesis was accomplished by measuring the blood vessel thickness. The number of blood vessel branches and the blood vessel thickness were counted in the pictures (4 pictures/egg) using Corel Draw 7. Thereafter, the number of blood vessel branches and the thickness of the blood vessels were correlated to a raster of microscope (10 mm<sup>2</sup>) for comparison.

25        The mean number of branches and the mean blood vessel thickness of each condition (n=10) were calculated and compared to the PBS control eggs using a Student's T-test.

## RESULTS

### Chicken eggs experiments

In order to determine the effect of NMPF on angiogenesis and vasculogenesis we treated fertilized chicken eggs with NMPF or NMPF in combination with VEGF as described in materials and methods section. Figures 20-30 show that NMPF 3, 4, 9 and 11 promoted angiogenesis ( $p < 0.05$ ), while NMPF VEGF, 7, 43, 44, 45, 46, 51 and 56 inhibited angiogenesis ( $p < 0.05$ ). NMPF 6, 7, 12, 45, 46 and 66 were able to inhibit angiogenesis induced by VEGF. Moreover, NMPF 6 itself did not show any effect on angiogenesis, but it inhibited ( $p < 0.05$ ) NMPF 3 induced angiogenesis. Figures 31-32 show that NMPF 1, 2, 3, 4, 6, 7, 8, 12, 50, 51, and 52 had vasculogenesis inhibiting ( $p < 0.05$ ) effect, while only NMPF 44 promoted ( $p < 0.05$ ) vasculogenesis.

### NOD experiment

*Mice.* Female NOD mice at the age of 13-14 weeks were treated i.p. with PBS (n=6) or NMPF peptides (VLPALPQVVC, LQGV, GVLPALPQ, VLPALP, VLPALPQ, MTRV, LPGCPRGVNPVVS, CPRGVNPVVS, LPGC, MTRVLQGVLPALPQVVC, VVCNYRDVRFESIRLPGCPRGVNPVVS, YAVALSQCQCAL) (5 mg/kg, n=6) three times a week for 2 weeks. Every four days urine was checked for the presence of glucose (Gluketur Test; Boehringer Mannheim, Mannheim, Germany). All mice used in these studies were maintained in a pathogen-free facility. They were given free access to food and water. The experiments were approved by the Animal Experiments Committee of the Erasmus University Rotterdam. Diabetes was assessed by measurement of the venous blood glucose level using an Abbott Medisense Precision glucometer. Mice were considered diabetic after two consecutive glucose measurements  $\geq 11$  mmol/l (200 mg/dl). Onset of diabetes was dated from the first consecutive reading.

Glucose tolerance test (GTT) test was performed at 28 weeks of age in fasted mice (n=5) by injecting 1 g/kg D-glucose intraperitoneally (i.p.). At 0 (fasting), 5, 30 and 60 minutes blood samples were collected from the tail and tested for glucose content.

### NO experiment

*Cell culture.* The RAW 264.7 murine macrophage cell line, obtained from American Type Culture Collection (Manassas, VA, USA), were cultured at 37°C in 5%



CO<sub>2</sub> using DMEM containing 10% fetal calf serum (FCS), 50 U/ml penicillin, 50 µg/ml streptomycin, 0.2 M Na-pyruvate, 2 mM glutamine and 50 µM 2-mercaptoethanol (Bio Whittaker, Europe). The medium was changed every 2 days.

*Nitrite measurements.* Nitrite production was measured in the RAW 264.7 macrophage supernatants. The cells ( $7.5 \times 10^5$ /ml) were cultured in 48-well plates in 500 µl of culture medium. The cells were stimulated with LPS (10 microg/ml) and/or NMPF (1 pg/ml, 1 ng/ml, 1 µg/ml) for 24 hours, then the culture media were collected. Nitrite was measured by adding 100 microl of Griess reagent (Sigma) to 100 microl samples of culture medium. The OD<sub>540</sub> was measured using a microplate reader, and the nitrite concentration was calculated by comparison with the OD<sub>540</sub> produced using standard solutions of sodium nitrite in the culture medium.

## RESULTS

### NOD experiment

In order to determine whether NMPF has effect on the disease development in NOD mice, we tested NMPF on pre-diabetic female NOD mice at the age of 13-14 weeks. After only two weeks of treatment (injection of NMPF (5 mg/kg) every other day) glucosuria data of all NOD mice was analyzed at the of 17 weeks. Profound anti-diabetic effect (mice negative for glucosuria) was observed in different NMPF groups as compared to PBS group, especially in NMPF groups treated with peptide VLPALPQVVC, VLPALP, MTRV, LPGCPRGVNPVVS and LPGC. In addition, impairment of the glucose tolerance test was positively correlated to insulinitis, but negatively correlated to the number of functional beta cells, also this test showed that NOD mice successfully treated with NMPF were tolerant for glucose as compared to PBS group. Our results show that PBS treated NOD mice were all diabetic at the age of 23 weeks. Whereas, NOD mice treated with three times a week for two weeks with NMPF showed profound inhibition of diabetes development. The strongest anti-diabetic effects were seen with NMPF-1, 4, 5, 6, 7, 65, 66 and commercial hCG preparation (Pregnyl, Organon, Oss, The Netherlands, batch no. 235863). These mice had a low fasting blood glucose level and were tolerant for glucose (data partially shown). However, NMPF-71 showed no effect on the incidence of diabetes, while NMPF-64 and NMPF-11 had a moderate anti-diabetic effect.

## NO experiment

NO production is a central mediator of the vascular and inflammatory response. Our results show that macrophages (RAW 264.7) stimulated with LPS produce large amount of NO. However, these cells co-stimulated with most of the NMPF peptides  
 5 (NMPF peptide 1 to 14, 43 to 66 and 69) even in a very low dose (1 pg/ml) inhibited the production of NO.

## Results

### ApoE experiment

10 The invention provides a method and a signaling molecule for the treatment of conditions that are associated with dysfunctional LDL receptors such as ApoE and other members of the apolipoprotein family. In particular, use of a signaling molecule comprising GVLPALPQ (NMPF – 5) and/or VLPALP (NMPF-6) or a functional analogue or derivative thereof is preferred. Groups of ApoE deficient mice (n=6 per group) were fed  
 15 a high cholesterol food and given PBS or NMPF every other day intraperitoneally. After 2.5 weeks body weight was determined as shown in the Table below.

	Average (g)	Weight SD (g)	p-value
ApoE-/- PBS	31.667	1.007	
ApoE-/- NMPF-43	31.256	1.496	0.536
ApoE-/- NMPF-52	29.743	1.160	0.019
Background/PBS	26.760	1.582	10 <sup>-6</sup>
ApoE-/- NMPF-62	29.614	1.064	0.004

### Analysis of different peptides in data bases

20 Examples of different data bases in which peptides were analysed are:

Proteomics tools: Similarity searches

BLAST data base (ExPasy, NCBI)

SMART (EMBL)

PATTINPROT (PBIL)

25 Post-translational modification prediction

SignalP (CBS)

Primary structure analysis

HLA Peptide Binding Predictions (BIMAS)

Prediction of MHC type I and II peptide binding

5 (SYFPEITHI)

Amino acid scale representation (Hydrophobicity, other conformational parameters, etc.) (PROTSKALE)

Representations of a protein fragment as a helical wheel(HelixWheel / HelixDraw)

10 RESULTS

A non-extensive list of relevant oligopeptides useful for application in a method to identify signaling molecules according to the invention derivable from protein data bases  
pdb|1DE7|1DE7-A INTERACTION OF FACTOR XIII ACTIVATION PEPTIDE WITH ALPHA- THROMBIN

15 LQGV, LQGVV, LQGVVP

pdb|1DL6|1DL6-A SOLUTION STRUCTURE OF HUMAN TFIIB N-TERMINAL DOMAIN

LDALP

pdb|1QMH|1QMH-A CRYSTAL STRUCTURE OF RNA 3'-TERMINAL PHOSPHATE

20 CYCLASE, AN UBIQUITOUS ENZYME

LQTV, VLPAL, LVLQTVLPAL

pdb|1LYP|1LYP CAP18 (RESIDUES 106 - 137)

IQG, IQGL, LPKL, LLPKL

pdb|1B9O|1B9O-A HUMAN ALPHA-LACTALBUMIN

25 LPEL

pdb|1GLU|1GLU-A GLUCOCORTICOID RECEPTOR (DNA-BINDING DOMAIN)

PARP

pdb|2KIN|2KIN-B KINESIN (MONOMERIC) FROM RATTUS NORVEGICUS

MTRI

30 pdb|1SMP|1SMP-I MOL\_ID: 1; MOLECULE: SERRATIA METALLO PROTEINASE;  
CHAIN: A

LQKL, LQKLL, PEAP, LQKLLPEAP

pdb|1ES7|1ES7-B COMPLEX BETWEEN BMP-2 AND TWO BMP RECEPTOR IA  
ECTODOMAINS

LPQ, PTLP, LQPTL

pdb|1BHX|1BHX-F X-RAY STRUCTURE OF THE COMPLEX OF HUMAN ALPHA  
5 THROMBIN WITH THE INHIBITOR SDZ 229-357

LQV, LQVV

pdb|1VCB|1VCB-A THE VHL-ELONGINC-ELONGINB STRUCTURE

PELP

pdb|1CQK|1CQK-A CRYSTAL STRUCTURE OF THE CH3 DOMAIN FROM THE  
10 MAK33 ANTIBODY

PAAP, PAAPQ, PAAPQV

pdb|1FCB|1FCB-A FLAVOCYTOCHROME

LQG,

pdb|1LDC|1LDC-A L-LACTATE DEHYDROGENASE: CYTOCHROME C  
15 OXIDOREDUCTASE (FLAVOCYTOCHROME B=2=) (E.C.1.1.2.3) MUTANT WITH  
TYR 143 REPLACED BY PHE (Y143F) COMPLEXED WITH PYRUVATE

LQG

pdb|1BFB|1BFB BASIC FIBROBLAST GROWTH FACTOR COMPLEXED WITH  
HEPARIN TETRAMER FRAGMENT

20 LPAL, PALP, PALPE

pdb|1MBF|1MBF MOUSE C-MYB DNA-BINDING DOMAIN REPEAT 1

LPN

pdb|1R2A|1R2A-A THE MOLECULAR BASIS FOR PROTEIN KINASE A

LQG, LTEL

25 pdb|1CKA|1CKA-B C-CRK (N-TERMINAL SH3 DOMAIN) (C-CRKSH3-N)  
COMPLEXED WITH C3G PEPTIDE (PRO-PRO-PRO-ALA-LEU-PRO-PRO-LYS-LYS-  
ARG)

PALP

pdb|1RLQ|1RLQ-R C-SRC (SH3 DOMAIN) COMPLEXED WITH THE PROLINE-  
30 RICH LIGAND RLP2 (RALPPLPRY) (NMR, MINIMIZED AVERAGE STRUCTURE)

LPPL, PPLP

- pdb|1TNT|1TNT MU TRANSPOSASE (DNA-BINDING DOMAIN) (NMR, 33  
STRUCTURES)  
LPG, LPGL, LPK
- 5 pdb|1GJS|1GJS-A SOLUTION STRUCTURE OF THE ALBUMIN BINDING DOMAIN  
OF STREPTOCOCCAL PROTEIN G  
LAAL, LAALP
- pdb|1GBR|1GBR-B GROWTH FACTOR RECEPTOR-BOUND PROTEIN 2 (GRB2, N-  
TERMINAL SH3 DOMAIN) COMPLEXED WITH SOS-A PEPTIDE (NMR, 29  
STRUCTURES)
- 10 LPKL, PKLP
- pdb|1A78|1A78-A COMPLEX OF TOAD OVARY GALECTIN WITH THIO-  
DIGALACTOSE  
VLPSIP
- 15 pdb|1ISA|1ISA-A IRON(II) SUPEROXIDE DISMUTASE (E.C.1.15.1.1)  
LPAL, PALP
- pdb|1FZV|1FZV-A THE CRYSTAL STRUCTURE OF HUMAN PLACENTA GROWTH  
FACTOR-1 (PLGF-1), AN ANGIOGENIC PROTEIN AT 2.0A RESOLUTION  
PAVP, MLPAPV
- 20 pdb|1JLI|1JLI HUMAN INTERLEUKIN 3 (IL-3) MUTANT WITH TRUNCATION AT  
BOTH N- AND C-TERMINI AND 14 RESIDUE CHANGES, NMR, MINIMIZED  
AVERAGE  
LPC, LPCL, PCLP
- pdb|1HSS|1HSS-A 0.19 ALPHA-AMYLASE INHIBITOR FROM WHEAT  
VPALP
- 25 pdb|3CRX|3CRX-A CRE RECOMBINASE/DNA COMPLEX INTERMEDIATE I  
LPA, LPAL, PALP
- pdb|1PRX|1PRX-A HORF6 A NOVEL HUMAN PEROXIDASE ENZYME  
PTIP, VLPTIP
- 30 pdb|1RCY|1RCY RUSTICYANIN (RC) FROM THIOBACILLUS FERROOXIDANS  
VLPGFP
- pdb|1A3Z|1A3Z REDUCED RUSTICYANIN AT 1.9 ANGSTROMS  
PGFP, VLPGFP

pdb|1GER|1GER-A GLUTATHIONE REDUCTASE (E.C.1.6.4.2) COMPLEXED WITH FAD

LPALP, PALP

pdb|1PBW|1PBW-A STRUCTURE OF BCR-HOMOLOGY (BH) DOMAIN

5 PALP

pdb|1BBS|1BBS RENIN (E.C.3.4.23.15)

MPALP

AI188872 11.3 366 327 18 382 [Homo sapiens]qd27c01.x1

Soares\_placenta\_8to9weeks\_2NbHP8to9W

10 Homo sapiens cDNA clone IMAGE:1724928 3' similar to gb:J00117 CHORIOGONADOTROPIN BETA CHAIN PRECURSOR (HUMAN);, mRNA sequence.; minus strand; translated

MXRVLQGVLPALPQVVC, MXRV, MXR,

15 AI126906 19.8 418 343 1 418 [Homo sapiens]qb95f01.x1 Soares\_fetal\_heart\_NbHH19W Homo sapiens cDNA clone IMAGE:1707865 3' similar to gb:J00117 CHORIOGONADOTROPIN BETA CHAIN PRECURSOR (HUMAN);, mRNA sequence.; minus strand; translated

ITRVMQGVIPALPQVVC

20

AI221581 29.1 456 341 23 510 [Homo sapiens]qg20a03.x1

Soares\_placenta\_8to9weeks\_2NbHP8to9W Homo sapiens cDNA clone IMAGE:1760044

3' similar to gb:J00117 CHORIOGONADOTROPIN BETA CHAIN PRECURSOR (HUMAN);, mRNA sequence.; minus strand; translated

25 MTRVLQVVLLALPQLV

Mm.42246.3 Mm.42246 101.3 837 304 28 768 GENE=Pck1 PROTSIM=pir:T24168 phosphoenolpyruvate carboxykinase 1, cytosolic; translated

30 KVIQGSLSLDPQAV, LDSL, LPQ

Mm.22430.1 Mm.22430 209.4 1275 157 75 1535 GENE=Ask-pending  
PROTSIM=pir:T02633 activator of S phase kinase; translated  
VLQAILPSAPQ, LQA, LQAIL, PSAP, LPS

5 Hs.63758.4 Hs.63758 93.8 3092 1210 51 2719 GENE=TFR2  
PROTSIM=pir:T30154 transferrin receptor 2; translated KVLQGRLPAVAQAV, LQG,  
LPA, LPAV

Mm.129320.2 Mm.129320 173.0 3220 571 55 2769 GENE= PROTSIM=pir:T16409  
10 Sequence 8 from Patent WO9950284; translated  
LVQKVVPMLPRLLC, LVQ, LPRL, PMLP

Mm.22430.1 Mm.22430 209.4 1275 157 75 1535 GENE=Ask-pending  
PROTSIM=pir:T02633 activator of S phase kinase; translated  
15 VLQAILPSAPQ, LQA, LQAIL, PSAP, PSAPQ

P20155 IAC2\_HUMAN Acrosin-trypsin inhibitor II precursor (HUSI-II) [SPINK2] [Homo  
sapiens]  
LPGCPRHFNPV, LPG, LPGC  
20

Rn.2337.1 Rn.2337 113.0 322 104 1 327 GENE= PROTSIM=PRF:1402234A Rat  
pancreatic secretory trypsin inhibitor type II (PSTI-II) mRNA, complete cds; minus strand;  
translated  
LVGCPRDYDPV, LVG, LVGC  
25

Hs.297775.1 Hs.297775 43.8 1167 753 31 1291 GENE= PROTSIM=sp:O00268 ESTs,  
Weakly similar to T2D3\_HUMAN TRANSCRIPTION INITIATION FACTOR TFIID 135  
KDA SUBUNIT [H.sapiens]; minus strand; translated  
PGCPRG, PGCP  
30

Mm.1359.1 Mm.1359 PROTSTM=pir.A39743 urokinase plasmiogen activator receptor  
LPGCP, PGCP, LPG, LPGC

sptrembl|O56177|O56177 ENVELOPE GLYCOPROTEIN  
VLPAAP, PAAP

5 sptrembl|Q9W234|Q9W234 CG13509 PROTEIN.//:trembl|AE003458|AE003458\_7 gene:  
"CG13509"; Drosophila melanogaster genomic scaffold  
LAGTIPATP, LAG, PATP

swiss|P81272|NS2B HUMAN NITRIC-OXIDE SYNTHASE IIB (EC 1.14.13.39) (NOS,  
10 TYPE II B) (NOSIIB) (FRAGMENTS)  
GVLPAVP, LPA, VLPAVP, PAVP

sptrembl|O30137|O30137 HYPOTHETICAL 17.2 KDA  
15 GVLPALP, PALP, LPAL

sptrembl|Q9IYZ3|Q9IYZ3 DNA POLYMERASE  
GLLPCLP, LPC, LPCL, PCLP

sptrembl|Q9PVW5|Q9PVW5 NUCLEAR PROTEIN NP220  
20 PGAP, LPQRPRGPNP, LPQ, PRGP, PNP

Hs.303116.2 PROTSIM=pir;T33097 stromal cell-derived factor 2-like1; translated  
GCPR

25 pdb|1DU3|1DU3-A CRYSTAL STRUCTURE OF TRAIL-SDR5  
GCPRGM

pdb|1D0G|1D0G-R CRYSTAL STRUCTURE OF DEATH RECEPTOR 5 (DR5) BOUND  
TO APO2L/TRAIL  
30 GCPRGM



pdb|1BIO|1BIO HUMAN COMPLEMENT FACTOR D IN COMPLEX WITH ISATOIC ANHYDRIDE INHIBITOR

LQHV

- 5 pdb|4NOS|4NOS-A HUMAN INDUCIBLE NITRIC OXIDE SYNTHASE WITH INHIBITOR

FPGC, PGCP

pdb|1FL7|1FL7-B HUMAN FOLLICLE STIMULATING HORMONE

PARP, VPGC

10

pdb|1HR6|1HR6-A YEAST MITOCHONDRIAL PROCESSING PEPTIDASE

CPRG, LKGC

- 15 pdb|1BFA|1BFA RECOMBINANT BIFUNCTIONAL HAGEMAN FACTOR/AMYLASE INHIBITOR FROM

PPGP, LPGCPREV, LPGC, PGCP, CPRE

- 20 swissnew|P01229|LSHB HUMAN Lutropin beta chain precursor

MMRVLQAVLPPLPQVVC, MMR, MMRV, LQA, LQAV, VLPPLP, PPLP, QVVC, VVC, VLPPLPQ, AVLPPPLP, AVLPPPLPQ

- 25 swissnew|P07434|CGHB PAPAN Choriogonadotropin beta chain precursor

MMRVLQAVLPVPQVVC, MMR, MMRV, LQA, LQAG, VLPPVP, VLPPVPQ, QVVC, VVC, AVLPPVP, AVLPPVPQ

- swissnew|Q28376|TSHB HORSE Thyrotropin beta chain precursor

MTRD, LPK, QDVC, DVC, IPGC, PGCP

- 30 swissnew|P95180|NUOB MYCTU NADH dehydrogenase I chain B

LPGC, PGCP

sptrembl|Q9Z284|Q9Z284 NEUTROPHIL ELASTASE

PALP, PALPS

sptrembl|Q9UCG8|Q9UCG8 URINARY GONADOTROPHIN PEPTIDE (FRAGMENT).

LPGGPR, LPG, LPGG, GGPR

5

XP\_028754 growth hormone releasing hormone [Homo sapiens]

LQRG, LQRGV, LGQL

SignalP (CBS)

10

SignalP predictions: (for example)

MTRVLQGVLPALP

QVVC

15

HLA Peptide Binding Predictions (BIMAS)

(For example)

Half time of dissociation

20	HLA molecule type I (A_0201):	VLQGVLPAL	(84)
		GVLPALQV	(51)
		VLPALPQVV	(48)
		RLPGCPRGV	(14)
		TMTRVLQGV	(115)

25

			scores
	MHC II (H2-Ak 15 – mers)	CPTMTRVLQGVLPAL	14
		PGCPRGVNPVVS YAV	14
	HLA-DRB1*0101 15 – mers	PRGVNPVVS YAVALS	29
30		TRVLQGVLPALPQVV	28
		LQGVLPALPQVVCNY	22
	HLA-DRB1*0301 (DR17)		

15 – mers

CPTMTRVLQGVLPAL	26
MTRVLQGVLPALPQV	21
SIRLPGCPRGVNPVV	17

5

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TABLE 1. Results of shock experiments in mice

TEST SUBSTANCE			% SURVIVAL IN TIME			
(HRS)			0	16	40	72
5	PBS		100	100	67	17
	PG23		100	100	100	100
	PG25		100	83	83	83
PEPTIDE						
10	NMPF	SEQUENCE				
	1	VLPALPQVVC	100	100	50	17
	2	LQGVLPALPQ	100	67	0	0
	3	LQG	100	83	20	17
15	4	LQGV	100	100	100	100
	5	GVLPALPQ	100	100	80	17
	6	VLPALP	100	100	100	100
	7	VLPALPQ	100	83	0	0
	8	GVLPALP	100	100	83	67
20	9	VVC	100	100	50	50
	11	MTRV	100	100	67	50
	12	MTR	100	100	67	50
	13	LQGVLPALPQVVC	100	100	100	100
	14	(CYCLIC) LQGVLPALPQVVC	100	83	83	83
25	64	LPGCPRGVNPVVS	100	100	100	100
	66	LPGC	100	100	100	100

TABLE 2. Additional results of shock experiments

NMPF SEQUENCE ID:		ANTI-SHOCK EFFECT
5	LQGV	+++
	AQGV	+++
	LQGA	+++
	VLPALP	+++
	ALPALP	++
10	VAPALP	++
	ALPALPQ	++
	VLPAAPQ	++
	VLPALAQ	+++
SHOCK ACCELERATING EFFECT		
15	LAGV	+++
	LQAV	+++
	VLAALP	+++
	VLPAAP	+++
	VLPALA	+++
20	VLPALPQ	+++
	VLAALPQ	+++
	VLPALPA	+++

TABLE 3. Further additional results of shock experiments

	NMPF PEPTIDES	% SURVIVAL IN TIME (HRS)			
		Tx	Tx		
5		0	14	24	48
	PBS	100	100	100	0
	NMPF-3	100	100	100	0
	NMPF-5	100	100	100	100
	NMPF-7	100	100	100	67
10	NMPF-8	100	100	100	100
	NMPF-9	100	100	100	100
	NMPF-11	100	100	100	100
	NMPF-12	100	100	100	100
	NMPF-43	100	100	100	100
15	NMPF-45	100	100	100	100
	NMPF-46	100	100	100	100
	NMPF-50	100	100	100	100
	NMPF-53	100	100	100	100
	NMPF-58	100	100	100	100
20	NMPF-60	100	100	100	100

TABLE 4. Further additional results

NMPF PEPTIDES		SICKNESS SCORES			
		Tx	Tx		
5		0	14	24	48
	PBS	0,0,0,0,0,0	5,5,5,5,4,4	5,5,5,5,5,5	††††††
	NMPF-3	0,0,0,0,0,0	3,3,3,3,3,4	4,4,4,4,4,4	††††††
	NMPF-5	0,0,0,0,0,0	5,5,5,5,5,5	5,5,5,5,5,5	2,2,2,2,2,2
10	NMPF-7	0,0,0,0,0,0	1,1,4,4,4,4	5,5,5,5,5,5	2,2,2,2,††
	NMPF-8	0,0,0,0,0,0	3,3,5,5,5,5	5,5,5,5,5,5	2,2,4,4,4,5
	NMPF-9	0,0,0,0,0,0	3,3,4,4,5,5	2,2,2,2,2,2	1,1,2,2,2,2
	NMPF-11	0,0,0,0,0,0	1,1,3,3,4,4	2,2,2,2,4,4	1,1,1,1,1,1
	NMPF-12	0,0,0,0,0,0	1,1,1,1,3,3	1,1,1,1,1,1	1,1,1,1,1,1
15	NMPF-43	0,0,0,0,0,0	1,1,4,4,4,4	1,1,1,1,3,3	2,2,2,2,2,2
	NMPF-45	0,0,0,0,0,0	5,5,5,5,4,4	3,3,4,4,5,5	2,2,4,4,5,5
	NMPF-46	0,0,0,0,0,0	1,1,2,2,3,3	1,1,2,2,2,2	1,1,1,1,1,1
	NMPF-50	0,0,0,0,0,0	1,1,1,1,3,3	2,2,2,2,3,3	1,1,1,1,1,1
	NMPF-53	0,0,0,0,0,0	5,5,5,5,5,5	5,5,5,5,5,5	1,1,2,2,2,2
20	NMPF-58	0,0,0,0,0,0	5,5,5,5,3,3	5,5,5,5,3,3	1,1,1,1,1,1
	NMPF-60	0,0,0,0,0,0	1,1,4,4,2,2	2,2,2,2,4,4	1,1,1,1,1,1

ID	SEQUENCE	SEPSIS	ANGIOGENSIS	CAO	DC	NOD
NMPF-1	VLPALPQVVC	-+		+	+	
NMPF-2	LQGVLPALPQ	-+			+	
NMPF-3	LQG	-+	+	+	+	
NMPF-4	LQGV	+	+	+	+	
NMPF-5	GVLPALPQ	-+			+	
NMPF-6	VLPALP	+	+	+	+	
NMPF-7	VLPALPQ	+	+		+	
NMPF-8	GVLPALP	-+			+	
NMPF-9	VVC	+	+		+	
NMPF-10	QVVC					
NMPF-11	MTRV	+	+		+	+
NMPF-12	MTR	-+	+		+	
NMPF-13	LQGVLPALPQVVC	+			+	
NMPF-14	cyclic- LQGVLPALPQVVC	+				
NMPF-43	AQG	+	+		+	
NMPF-44	LAG		+			
NMPF-45	LQA	+	+			
NMPF-46	AQGV	+	+		+	
NMPF-47	LAGV	-+		+	+	
NMPF-48	LQAV					
NMPF-49	LQGA	+				
NMPF-50	ALPALP	+			+	
NMPF-51	VAPALP	+	+			
NMPF-52	VLAALP					
NMPF-53	VLPAAP	+			+	
NMPF-54	VLPALA					
NMPF-55	ALPALPQ	+				
NMPF-56	VAPALPQ		+			
NMPF-57	VLAALPQ					
NMPF-58	VLPAAPQ	+			+	
NMPF-59	VLPALAQ	+	+			
NMPF-60	VLPALPA	+			+	
NMPF-61	VVCNRYRDVRFESIRLPGCPRGVNPVVSYAVALSCQCAL	-+		+		
NMPF-62	VVCNRYRDVRFESIRLPGCPRGVNPVVSYAVALSCQ					
NMPF-63	SIRLPGCPRGVNPVVS	-+				
NMPF-64	LPGCPRGVNPVVS			+		
NMPF-65	CPRGVNPVVS					
NMPF-66	LPGC	+	+	+		
NMPF-67	CPRGVNP					
NMPF-68	PGCP	-+				
NMPF-69	RPRCRPINATLAVEKEGCPVCITVNTTICAGYCPT					
NMPF-70	MTRVLQGVLPALPQ	-+				
NMPF-71	MTRVLPGVLPALPQVVC	-+				
NMPF-74	CALCRRSTTDCGGPKDHPLTC					
NMPF-75	SKAPPPSLPSPSRLPGPC					
NMPF-76	TCDDPRFQDSSSSKAPPPSLPSPSRLPGPSDTPIPLQ					

Table 5 Summary of results of the various peptides in the various experiments.

+ = effects; -+ = variable effect; no entry is no effect or not yet tested when table was

5 assembled



**MODULATION OF NO AND/OR**

**Table 6 TNF-ALPHA**

<b>ID</b>	<b>SEQUENCE</b>	<b>TNF-A</b>	<b>NO</b>	<b>TNF-A and NO</b>
<b>NMPF-1</b>	VLPALPQVVC	++	++++	++++
<b>NMPF-2</b>	LQGVLPALPQ	-+	++++	++++
<b>NMPF-3</b>	LQG	+	++++	++++
<b>NMPF-4</b>	LQGV	++++	++++	++++++
<b>NMPF-5</b>	GVLPALPQ	++++	++++	++++++
<b>NMPF-6</b>	VLPALP	++++	++++	++++++
<b>NMPF-7</b>	VLPALPQ	++++	++++	++++++
<b>NMPF-8</b>	GVLPALP	++++	++++	++++++
<b>NMPF-9</b>	VVC	++++	++++	++++++
<b>NMPF-10</b>	QVVC	++++	+++	++++
<b>NMPF-11</b>	MTRV	++++	++++	++++
<b>NMPF-12</b>	MTR	++++	++++	++++
<b>NMPF-13</b>	LQGVLPALPQVVC	++	++++	++++
<b>NMPF-14</b>	cyclic- LQGVLPALPQVVC	++	++++	++++
<b>NMPF-43</b>	AQG	++++	++++	++++++
<b>NMPF-44</b>	LAG	-+	++++	++++
<b>NMPF-45</b>	LQA	++++	++++	++++++
<b>NMPF-46</b>	AQGV	++++	++++	++++++
<b>NMPF-47</b>	LAGV	++	++++	++++
<b>NMPF-48</b>	LQAV	++	++++	++++
<b>NMPF-49</b>	LQGA	++	++++	++++
<b>NMPF-50</b>	ALPALP	++++	++++	++++++
<b>NMPF-51</b>	VAPALP	+	+++	++++
<b>NMPF-52</b>	VLAALP	++	++++	++++
<b>NMPF-53</b>	VLPAAP	++++	++++	++++++
<b>NMPF-54</b>	VLPALA	+	++++	++++

NMPF-55	ALPALPQ	+	++++	++++
<b>MODULATION OF NO AND/OR</b>				
<b>TNF-ALPHA</b>				
Table 6 cnt.				
ID	SEQUENCE	TNF-A	NO	TNF-A and NO
NMPF-56	VAPALPQ	-+	++++	++++
NMPF-57	VLAALPQ	+	++++	++++
NMPF-58	VLPAAPQ	++++	++++	++++++
NMPF-59	VLPALAQ	++	++++	++++
NMPF-60	VLPALPA	++++	++++	++++++
NMPF-61	VVCNYRDVRFESIRLPGCPRGVN			
	PVVSYAVALSCQCAL	-+	++++	++++
NMPF-62	VVCNYRDVRFESIRLPGCPRGVN			
	PVVSYAVALSCQ	-+	+++	++++
NMPF-63	SIRLPGCPRGVNPVVS	-+	++	++
NMPF-64	LPGCPRGVNPVVS	++	++++	++++
NMPF-65	CPRGVNPVVS	++	+++	+++
NMPF-66	LPGC	+++	++	+++
NMPF-67	CPRGVNP	-+	+	+
NMPF-68	PGCP	+	+	+++
NMPF-69	RPRCRPINATLAVEK			
	EGCPVCITVNTTICAGYCPT	-+	++	++
NMPF-70	MTRVLQGVLPALPQ	-+	+	+
NMPF-71	MTRVLPGVLPALPQVVC	-+	-+	-+
NMPF-74	CALCRRSTTDCGGPKDHPLTC	-+	++	+
NMPF-75	SKAPPPSLPSPSRLPGPS	+	++	++
NMPF-76	TCDDPRFQDSSSSKAPPPSLPSPSR			
	LPGPSDTPILPQ	+	+	+
NMPF-78	CRRSTTDCGGPKDHPLTC	+	+	+

from -+ to ++++++ indicates from barely active to very active in modulating

## Monkey experiment

Efficacy of NMPF, here a mixture 1:1:1 of LQGV, AQGV and VLPALP, administered in a gram-negative induced rhesus monkey sepsis model for prevention of septic shock.

- 5           Overwhelming inflammatory and immune responses are essential features of septic shock and play a central part in the pathogenesis of tissue damage, multiple organ failure, and death induced by sepsis. Cytokines, especially tumor necrosis factor (TNF)- $\alpha$  interleukin (IL)-1 $\alpha$ , and macrophage migration inhibitory factor (MIF) have been shown to be critical mediators of septic shock. Yet, traditional anti-TNF and anti-IL-1 therapies
- 10       have not demonstrated much benefit for patients with severe sepsis. We have designed peptides that block completely LPS induced septic shock in mice, even when treatment with these peptides is started up to 24 hours after LPS injection. These peptides are also able to inhibit the production of MIF. This finding provides the possibility of therapeutic use of these peptides for the treatment of patients suffering from septic shock. Since
- 15       primates are evolutionary more closer to humans, we tested these peptides for their safety and effectiveness in a primate system.

# EXPERIMENTAL DESIGN

GROUP	EXPERIMENTAL TREATMENT (independent variable, e.g., placebo treated control group)	BIOTECHNIQUES	NUMBER	
animal I	i.v. infusion of a lethal dose of live Escherichia.coli (10E10 CFU/kg) + antibiotics + placebo treated	Live E. coli infusion Blood sampling No recovery (section)	N=1	
animal II	i.v. infusion of a lethal dose of live Escherichia.coli (10E10 CFU/kg) + antibiotics + oligopeptide ( 5mg/kg of each of 3 peptides)	Live E. coli infusion Blood sampling No recovery (section)	N=1	

- 5 Only naive monkeys were used in this pre-clinical study to exclude any interaction with previous treatments. The animals were sedated with ketamine hydrochloride. Animals were intubated orally and are allowed to breathe freely. The animals were kept anesthetized with O<sub>2</sub>/N<sub>2</sub>O/isoflurane. The animals received atropin as pre-medication for O<sub>2</sub>/N<sub>2</sub>O/isoflurane anesthesia. A level of surgical anesthesia was maintained during the 2
- 10 h infusion of *E. coli* and for 6 h following *E. coli* challenge after which the endotracheal tubes were removed and the animals were euthanized. Before bacteria were induced, a 1 hour pre-infusion monitoring of heart-rate and blood pressure was performed.

Two rhesus monkeys were infused with a  $10^{10}$  CFU per kg of the Gram negative bacterium *E. coli* to induce a fatal septic shock. One monkey received placebo-treatment and was sacrificed within 7 hours after infusion of the bacteria without recovery from the anesthesia. The second monkey received treatment with test compound and was sacrificed  
5 at the same time point.

In a limited dose-titration experiment performed with the same bacterium strain in 1991, the used dose proved to induce fatal shock within 8 hours. In recent experiments a 3-fold lower dose was used inducing clear clinical and pathomorphological signs of septic shock without fatal outcome.

10 The monkeys were kept anaesthetized throughout the observation period and sacrificed 7 hours after the start of the bacterium infusion for pathological examination. The animals underwent a gross necropsy in which the abdominal and thoracic cavities were opened and internal organs examined in situ.

#### 15 Full description of the experiment with three rhesus monkeys

The study was conducted in rhesus monkeys (*Macaca mulatta*). Only experimentally naïve monkeys were used in study to exclude any interaction with previous treatments. Prior to the experiment the state of health of the animals was assessed physically by a veterinarian. All animals had been declared to be in good health and were  
20 free of pathogenic ecto- and endoparasites and common bacteriological infections: *Yersinia pestis*, *Yersinia enterocolitica*, *Yersinia pseudotuberculosis*, *Shigella*, *Aeromonas hydrophila*, pathogenic *Campylobacter* species and *Salmonella*.

*Reagents.* The *Escherichia coli* strain was purchased from ATCC (*E. coli*; 086a: K61 serotype, ATCC 33985). In a control experiment the strain proved equally susceptible  
25 to bactericidal factors in human and rhesus monkey serum. Prior to the experiment a fresh culture was set-up; the *E. coli* strain was cultured for one day, harvested and washed thoroughly to remove free endotoxin. Prior to infusion into the animal the number and viability of the bacteria was assessed. Serial dilutions of the *E. coli* stock were plated on BHI agar and cultured overnight at 37°C. The colonies on each plate were counted and the  
30 number of colony forming units per ml were calculated. The body weight measurement of the day of the experiment was used to calculate the *E. coli* dose and *E. coli* stock was suspended in isotonic saline (N.P.B.I., Emmer-Compascuum, The Netherlands) at the

concentration needed for infusion (total dose volume for infusion approximately 10 ml/kg. The E. coli suspension was kept on ice until infusion. Antibiotic was used to synchronise the shock induction in the monkeys. Baytril (Baytril 2.5%, Bayer, Germany) was used instead of gentamycin, as the strain proved only marginally susceptible to the latter  
5 antibiotic. Individual animals were identified by a number or letter combination tattooed on the chest.

*Experimental design.*

GROUP (number/ letter or other identification)	EXPERIMENTAL TREATMENT (independent variable, e.g., placebo treated control group)		NUMBER	SEX
Animal I	i.v. infusion of a lethal dose of live Escherichia.coli (10E10 CFU/kg) + antibiotic + placebo treated	Live E. coli infusion Blood sampling No recovery	N=1	F
Animal II	i.v. infusion of a lethal dose of live Escherichia.coli (10E10 CFU/kg) + antibiotic + NMPF-4, 6, 46; each 5mg/kg	Live E. coli infusion Blood sampling No recovery (section)	N=1	F
Animal III	i.v. infusion of a lethal dose of live Escherichia.coli (10E10 CFU/kg) + antibioti + NMPF-4, 6, 46; each 5mg/kg	Live E. coli infusion Blood sampling Recovery and survival	N=1	F

*Anesthesia.* All animals were fasted overnight prior to the experiment. On the morning of the experiment the animals were sedated with ketamine hydrochloride (Tesink, The Netherlands) and transported to the surgery. The animal was placed on its side on a temperature controlled heating pad to support body temperature. Rectal temperature was monitored using a Vet-OX 5700. The animals were intubated orally and were allowed to breathe freely. The animals were kept anesthetized using O<sub>2</sub>/N<sub>2</sub>O/isoflurane inhalation anaesthesia during the *E. coli* infusion and the 7 hour observation period following *E. coli* challenge after which the endotracheal tubes were removed and the animals were euthanized or allowed to recover from anesthesia. The femoral or the cephalic vein was cannulated and used for infusing isotonic saline, live *E. coli* and antibiotic administration. Insensible fluid loss was compensated for by infusing isotonic saline containing 2.5% glucose (Fresenius, 's Hertogenbosch, NL) at a rate of 3.3 ml/kg/hr.

*Preparative actions.* During anesthesia the animals were instrumented for measurement of blood pressure (with an automatic cuff), heart rate and body temperature. Isotonic saline was infused at 3.3 ml/kg/hr to compensate for fluid loss. Femoral vessels were cannulated for infusion of *E. coli* and antibiotics. Temperature controlled heating pads were used to support body temperature. The monkeys were continuously monitored during *E. coli* challenge and for the 6 hr period following *E. coli* administration. After 7 hrs 2 animals (the control animal and one treated with NMPF) were sacrificed to compare the direct effect of the compound at the level of histology. The 3<sup>rd</sup> animal, treated with NMPF, was allowed to recover from anaesthesia and was intensively observed during the first 12 hours after recovery followed by frequent daily observation. The decision to allow 3<sup>rd</sup> animal to recover was made after consulting with the veterinarian.

*Induction of septic shock.* Before the infusion of *E. coli*, a 1 hr pre-infusion monitoring of heart-rate and blood pressure was performed. All three animals received an i.v. injection of *E. coli* 086 (k61 serotype; ATCC 33985) at a lethal dose of 10 x10<sup>9</sup> CFU/kg bodyweight. In a dose titration study with this batch performed in 1991, this bacterial dose induced lethal shock within 8 hrs after the start of the infusion. The infusion period was 2 hrs.

*Antibiotics.* Baytril was administered intravenously immediately after completion of the 2 h. *E. coli* infusion (i.v.; dose 9 mg/kg).



*Treatment with NMPF.* 30 minutes post-onset of *E. coli* infusion the animals were administered a single intravenous bolus injection a mixer of NMPF oligopeptide. The oligopeptide mixer contained the following NMPF peptides: LQGV (5 mg/kg), AQGV (5 mg/kg) and VLPALP (5 mg/kg). These NMPF peptides were dissolved in 0.9% sodium chloride for injection (N.P.B.I., Emmer Compascuum, The Netherlands).

## RESULTS

### Preliminary monkey results

An anti-shock effect of the test compound on sepsis in the monkey treated with the oligopeptide mixture, namely the inhibition of the effect of the sepsis in this early 7-hour trajectory of this primate model was observed. Immunomodulatory effects with these peptides have been observed *in vitro/ex vivo* such as in T-cell assays the inhibition of pathological Th1 immune responses, suppression of inflammatory cytokines (MIF), increase in production of anti-inflammatory cytokines (IL-10, TGF-beta) and immunomodulatory effects on antigen presenting cells (APC) like dendritic cells and macrophages.

The following organs were weighed and a bacterial count were performed: kidneys, liver, lungs, lymph nodes, and gross lesions

Tissues of all organs were preserved in neutral aqueous phosphate buffered 4% solution of formaldehyde. Lymphoid organs were be cryopreserved. All tissues will be processed for histopathological examination.

### Further results obtained in the three-monkey experiment

*Monkey 429(control).* Female monkey (5.66 kg) received an i.v. injection of *E. coli* 086 (10E10 CFU/kg). In a dose titration study with this batch performed in 1991, this bacterial dose induced lethal shock within 8 hrs after the start of the infusion. The infusion period was 2 hrs. Baytril was administered intravenously immediately after completion of the 2 h. *E. coli* infusion (i.v.; dose 9 mg/kg). After the *E. coli* injection the monkey was observed by the authorized veterinarian without knowing which of the monkey received NMPF treatment. The clinical observations were as follows: vomiting, undetectable pulse, heart arrhythmia, abnormalities in ECG: signs of ventricle dilatation/heart decompensation (prolonged QRS complex, extra systoles), decreased blood clotting and forced respiration.

In addition, there was big fluctuation in heart rate (30-150 beats per minute), collapse of both systolic and diastolic blood pressure (35/20 mmHg) and decrease in blood oxygen concentration (80-70%). Seven hours after the start of the *E. coli* infusion, monkey began to vomit blood and faeces, and have convulsions. After final examination, the veterinarian did not give permission to let this monkey awake. At this time point control monkey was euthanised. Hereafter, post mortem examination was conducted and internal organs were examined in situ. Number of internal bleedings were found by the pathologist.

*Monkey 459(NMPF)*. Female monkey (5.44 kg) received an i.v. injection of *E. coli* 086 (10E10 CFU/kg). In a dose titration study with this batch performed in 1991, this bacterial dose induced lethal shock within 8 hrs after the start of the infusion. The infusion period was 2 hrs. 30 minutes after the initiation of *E. coli* infusion, NMPF was i.v. injected in a single bolus injection. Baytril was administered intravenously immediately after completion of the 2 h. *E. coli* infusion (i.v.; dose 9 mg/kg). After the *E. coli* injection this monkey was also observed by the authorized veterinarian without knowing which of the monkey received NMPF treatment. The clinical observations were as follows: normal pulse, heart sounds normal, normal ECG, higher heart-rate but otherwise stable (180 beats per minute), no hypotension (75/30 mmHg), normal blood oxygen concentration (95-85%), lungs sound normal, normal turgor. Seven hours after the start of the *E. coli* infusion, the clinical condition of the monkey was stable. After final examination, the veterinarian did give permission to let this monkey awake due to her stable condition. In order to compare the hematological and immunological parameters between the control and NMPF treated monkey, at this time point NMPF treated monkey 459 was euthanised. Hereafter, post mortem examination was conducted and internal organs were examined in situ. No macroscopic internal bleedings were found by the pathologist.

*Monkey 427(NMPF)*. Female monkey (4.84 kg) received an i.v. injection of *E. coli* 086 (10E10 CFU/kg). In a dose titration study with this batch performed in 1991, this bacterial dose induced lethal shock within 8 hrs after the start of the infusion. The infusion period was 2 hrs. 30 minutes after the initiation of *E. coli* infusion, NMPF was i.v. injected. Baytril was administered intravenously immediately after completion of the 2 h. *E. coli* infusion (i.v.; dose 9 mg/kg). After the *E. coli* injection this monkey was also observed by the authorised veterinarian doctor without knowing which of the monkey received NMPF treatment. The clinical observations were as follows: normal pulse, heart sounds normal,

normal ECG, moderately higher heart-rate but otherwise stable (160 beats per minute), no hypotension (70/30 mmHg), normal blood oxygen concentration (95-90%), lungs sound normal, normal turgor. Seven hours after the start of the *E. coli* infusion, the clinical condition of the monkey was stable. After final examination, the veterinarian did give permission to let this monkey wake up due to her stable condition. Monkey woke up quickly, she was alert and there was a slow disappearance of oedema.

### Genomic experiment

PM1 T-cell line was obtained from American Type Culture Collection (Manassas, VA) and was cultured at 37°C in 5% CO<sub>2</sub>. These cells were maintained and cultured in RPMI 1640, 10% fetal bovine serum, 2 mM L-glutamine, and antibiotics penicillin and streptomycin. For genomic experiments cells ( $2 \times 10^6$ /ml) were incubated with phytohemagglutinin (PHA, 10 µg/ml) and IL-2 (200 IU/ml) or PHA, IL-2 and peptide LQGV (10mg/ml) in a volume of 2 ml in 6-wells plates. After 4 h of cultures  $10 \times 10^6$  cells were washed and prepared for genechip probe arrays experiment. The genechip expression analysis was performed according to the manufacturer's instructions (Expression Analysis, Technical Manual, Affymetrix Genechip). The following major steps outline Genechip Expression Analysis: 1) Target preparation 2) Target hybridization 3) Experiment and fluidics station setup 4) Probe Array washing and staining 5) Probe array scan and 6) Data analysis.

## RESULTS

### Genomic experiment

The gene chip expression analysis revealed that due to LQGV treatment of PM1 (T-cell line) cells for 4 hours in the presence of PHA/IL-2 down-regulated at least 120 genes more than 2 fold as compared to control PM1 cells (stimulated with PHA/IL-2) only. Moreover, at least 6 genes were up-regulated more than 2 fold in peptide treated cells as compared to control cells.

Down regulated genes due to treatment with LQGV in genomics experiment

Fold Change/Descriptions

	21.2	M11507 Human transferrin receptor mRNA, complete cds (_5, _M, _3 represent transcript regions 5 prime, Middle, and 3 prime respectively)
5		
	10.1	Human (c-myb) gene, complete primary cds, and five complete alternatively spliced cds (U22376 /FEATURE=cds#5 /DEFINITION=HSU22376)
10		
	9.7	Cluster Incl. X68836:H.sapiens mRNA for S-adenosylmethionine synthetase (cds=(65,1252) /gb=X68836 /gi=36326 /ug=Hs.77502 /len=1283)
15	9.3	M97935 Homo sapiens transcription factor ISGF-3 mRNA, complete cds (_5, _MA, MB, _3 represent transcript regions 5 prime, MiddleA, MiddleB, and 3 prime respectively)
	8.7	Human mRNA for phosphatidylinositol transfer protein (PI-TPbeta), complete cds (D30037 /FEATURE= /DEFINITION=HUMPITPB)
20		
	7.5	Cluster Incl. U28964:Homo sapiens 14-3-3 protein mRNA, complete cds (cds=(126,863) /gb=U28964 /gi=899458 /ug=Hs.75103 /len=1030)
25	6.7	Human CDK tyrosine 15-kinase WEE1Hu (Wee1Hu) mRNA, complete cds (U10564 /FEATURE= /DEFINITION=HSU10564)
	6.7	Homo sapiens E2F-related transcription factor (DP-1) mRNA, complete cds (L23959 /FEATURE= /DEFINITION=HUMDP1A)
30		

- 6.5 Cluster Incl. W29030:55c4 Homo sapiens cDNA  
(gb=W29030 /gi=1308987  
/ug=Hs.4963 /len=758)
- 5 6.1 Cluster Incl. U08997:Human glutamate dehydrogenase gene, complete cds  
(cds=(0,1676) /gb=U08997 /gi=478987 /ug=Hs.239377 /len=1677)
- 5.7 M97935 Homo sapiens transcription factor ISGF-3 mRNA, complete cds  
(\_5, \_MA, MB, \_3 represent  
10 transcript regions 5 prime, MiddleA, MiddleB, and 3 prime respectively)
- 5.6 Cluster Incl. Y00638:Human mRNA for leukocyte common antigen (T200)  
(cds=(86,4000) /gb=Y00638 /gi=34280 /ug=Hs.170121 /len=4315)
- 15 5.3 Ras-Like Protein Tc21
- 5.3 H.sapiens mRNA for Fas/Apo-1 (clone pCRTM11-Fasdelta(4,7))  
( X83492  
/FEATURE=exons#1-2 /DEFINITION=HSFAS47)
- 20 4.8 Cluster Incl. AJ002428:Homo sapiens VDAC1 pseudogene  
(cds=(0,853) /gb=AJ002428 /gi=3183956 /ug=Hs.201553 /len=854)
- 25 4.7 Ras-Related Protein Rap1b
- 4.6 Cluster Incl. AL080119:Homo sapiens mRNA; cDNA DKFZp564M2423  
(from clone DKFZp564M2423) (cds=(85,1248) /gb=AL080119  
/gi=5262550 /ug=Hs.165998 /len=2183)
- 30 4.5 Cluster Incl. AF047448:Homo sapiens TLS-associated protein TASR  
mRNA, complete cds

(cds=(29,580) /gb=AF047448 /gi=2961148 /ug=Hs.239041 /len=620)

4.5 Cluster Incl. D14710:Human mRNA for ATP synthase alpha subunit,  
complete cds

5 (cds=(63,1724) /gb=D14710 /gi=559324 /ug=Hs.155101 /len=1857)

4.5 Cluster Incl. X59618:H.sapiens RR2 mRNA for small subunit  
ribonucleotide reductase

10 (cds=(194,1363) /gb=X59618 /gi=36154 /ug=Hs.75319 /len=2475)

4.5 Human mRNA for annexin II, 5' UTR (sequence from the 5' cap to the start  
codon)

(D28364 /FEATURE= /DEFINITION=HUMAI23)

15 4.5 Cluster Incl. AA477898:zu34f08.r1 Homo sapiens cDNA, 5' end  
/clone=IMAGE-739911 /clone\_end=5

(gb=AA477898 /gi=2206532 /ug=Hs.239414 /len=449)

4.4 Cluster Incl. L19161:Human translation initiation factor eIF-2 gamma  
20 subunit mRNA, complete cds (cds=(0,1418) /gb=L19161  
/gi=306899 /ug=Hs.211539 /len=1440 gb=AA477898 /  
gi=2206532 /ug=Hs.239414 /len=449)

4.4 Human serine/threonine-protein kinase PRP4h (PRP4h) mRNA, complete  
25 cds

(U48736 /FEATURE= /DEFINITION=HSU48736)

4.4 Cluster Incl. L43821:Homo sapiens enhancer of filamentation (HEF1)  
mRNA, complete cds  
30 (cds=(163,2667) /gb=L43821 /gi=1294780 /ug=Hs.80261 /len=3817)

- 4.4 Ras-Like Protein Tc21
- 4.4 Human (c-myb) gene, complete primary cds, and five complete alternatively  
spliced cds (U22376 /FEATURE=cds#3  
5 /DEFINITION=HSU22376)
- 4.3 Cluster Incl. U18271:Human thymopoietin (TMPO) gene  
(cds=(313,2397) /gb=U18271  
/gi=2182141 /ug=Hs.170225 /len=2796)
- 10 4.2 Fk506-Binding Protein, Alt. Splice 2
- 4.2 Human proliferating cell nuclear antigen (PCNA) gene, promoter region  
( J05614 /FEATURE=mRNA  
/DEFINITION=HUMPCNAPRM)
- 15 4.1 Human insulin-stimulated protein kinase 1 (ISPK-1) mRNA, complete cds  
(U08316 /FEATURE=  
/DEFINITION=HSU08316)
- 20 4.1 Cluster Incl. W28732:50h7 Homo sapiens cDNA  
(gb=W28732 /gi=1308680  
/ug=Hs.177496 /len=818)
- 4.1 Cluster Incl. Y00638:Human mRNA for leukocyte common antigen (T200)  
(cds=(86,4000) /gb=Y00638  
25 /gi=34280 /ug=Hs.170121 /len=4315)
- 4 Homo sapiens putative purinergic receptor P2Y10 gene, complete cds  
(AF000545 /FEATURE=cds  
/DEFINITION=HSAF000545)
- 30 3.8 Cluster Incl. U08997:Human glutamate dehydrogenase gene, complete cds  
(cds=(0,1676) /gb=U08997

/gi=478987 /ug=Hs.239377 /len=1677)

- 3.6 Human mRNA for raf oncogene
- 5 (X03484 /FEATURE=cds /DEFINITION=HSRAFR)
- 3.6 Cluster Incl. M32886:Human sorcin CP-22 mRNA, complete cds  
(cds=(12,608) /gb=M32886  
/gi=338481 /ug=Hs.117816 /len=952)
- 10 3.6 Homo sapiens GTP-binding protein (RAB1) mRNA, complete cds  
(M28209 /FEATURE= /DEFINITION=HUMRAB1A)
- 3.5 Human FKBP-rapamycin associated protein (FRAP) mRNA, complete cds
- 15 (L34075 /FEATURE= /DEFINITION=HUMFRAPX)
- 3.5 Human DNA topoisomerase II (top2) mRNA, complete cds
- 20 (J04088 /FEATURE= /DEFINITION=HUMTOPII)
- 3.4 Human translation initiation factor eIF-2 gamma subunit mRNA, complete  
cds  
(L19161 /FEATURE= /DEFINITION=HUMIEF2G)
- 25 3.4 Human mRNA for pre-mRNA splicing factor SRp20, 5' UTR (sequence  
from the 5' cap to the start codon)  
(D28423 /FEATURE= /DEFINITION=HUMPSF82)
- 30 3.4 Cluster Incl. AA442560:zv75g07.r1 Homo sapiens cDNA, 5' end  
/clone=IMAGE-759516 /clone\_end=5



(gb=AA442560 /gi=2154438 /ug=Hs.135198 /len=566)

3.4 Cluster Incl. X98248:H.sapiens mRNA for sortilin /cds=(21,2522)

5 (gb=X98248 /gi=1834494 /ug=Hs.104247 /len=3723)

3.3 Cluster Incl. AB020670:Homo sapiens mRNA for KIAA0863 protein,  
complete cds

10 (cds=(185,3580) /gb=AB020670 /gi=4240214 /ug=Hs.131915 /len=4313)

3.3 Cluster Incl. W28869:53h2 Homo sapiens cDNA

(gb=W28869  
/gi=1308880 /ug=Hs.74637 /len=975)

15 3.3 Cluster Incl. Z12830:H.sapiens mRNA for SSR alpha subunit /cds=(29,889)  
(gb=Z12830 /gi=551637 /ug=Hs.76152 /len=974)

3.3 Cluster Incl. AL021546:Human DNA sequence from BAC 15E1 on  
20 chromosome 12.  
Contains Cytochrome C Oxidase Polypeptide VIa-liver precursor gene,  
60S ribosomal protein L31 pseudogene, pre-mRNA splicing factor SRp30c  
gene,

25 two putative genes, ESTs, STSs and putative CpG islands  
(cds=(0,230) /gb=AL021546  
/gi=2826890 /ug=Hs.234768 /len=547)

3.2 Cluster Incl. U78082:Human RNA polymerase transcriptional regulation  
mediator (h-MED6) mRNA, complete cds

30 (cds=(50,523) /gb=U78082 /gi=2618737 /ug=Hs.167738 /len=885)

- 3.2 H.sapiens RbAp48 mRNA encoding retinoblastoma binding protein  
(X74262 /FEATURE=cds /DEFINITION=HSRBAP48)
- 5 3.1 Cluster Incl. M64174:Human protein-tyrosine kinase (JAK1) mRNA,  
complete cds  
(cds=(75,3503) /gb=M64174 /gi=190734 /ug=Hs.50651 /len=3541)
- 10 3.1 Cluster Incl. AI862521:wj15a06.x1 Homo sapiens cDNA, 3 end  
/clone=IMAGE-2402866 /clone\_end=3  
(gb=AI862521 /gi=5526628 /ug=Hs.146861 /len=606)
- 15 3.1 Cluster Incl. W27517:31h6 Homo sapiens cDNA  
(gb=W27517 /gi=1307321 /ug=Hs.13662 /len=732)
- 3 Human rab GDI mRNA, complete cds  
(D13988 /FEATURE= /DEFINITION=HUMRABGDI)
- 20 3 Cluster Incl. AL080119:Homo sapiens mRNA; cDNA DKFZp564M2423  
(from clone DKFZp564M2423)  
(cds=(85,1248) /gb=AL080119 /gi=5262550 /ug=Hs.165998 /len=2183)
- 25 3 Human cAMP-dependent protein kinase type I-alpha subunit (PRKAR1A)  
mRNA, complete cds  
( M33336 /FEATURE= /DEFINITION=HUMCAMPPK)
- 30 3 Cluster Incl. L75847:Human zinc finger protein 45 (ZNF45) mRNA,  
complete cds (cds=(103,2151) /gb=L75847  
/gi=1480436 /ug=Hs.41728 /len=2409)

- 3 Cluster Incl. M21154:Human S-adenosylmethionine decarboxylase mRNA,  
complete cds  
(cds=(248,1252) /gb=M21154 /gi=178517 /ug=Hs.75744 /len=1805)
- 5 3 Cluster Incl. AA675900:g02504r Homo sapiens cDNA, 5 end  
/clone=g02504 /clone\_end=5  
(gb=AA675900 /gi=2775247 /ug=Hs.119325 /len=647)
- 10 3 Cluster Incl. M97936:Human transcription factor ISGF-3 mRNA sequence  
(cds=UNKNOWN /gb=M97936 /gi=475254 /ug=Hs.21486 /len=2607)
- 2 M33336 /DEFINITION=HUMCAMPPK Human cAMP-dependent protein kinase  
type I-alpha subunit (PRKAR1A)  
mRNA, complete cds
- 15 2 U16720 /FEATURE=mRNA /DEFINITION=HSU16720 Human interleukin 10  
(IL10) gene, complete cds
- 2 M33336 HUMCAMPPK Human cAMP-dependent protein kinase type I-alpha  
subunit (PRKAR1A) mRNA
- 2 U50079 /FEATURE= /DEFINITION=HSU50079 Human histone deacetylase HD1  
mRNA, complete cds
- 20 2 U16720 /FEATURE=mRNA /DEFINITION=HSU16720 Human interleukin 10  
(IL10) gene, complete cds
- 2 X87212 /FEATURE=cds /DEFINITION=HSCATHCGE H.sapiens mRNA for  
cathepsin C
- 25 2 Cluster Incl. AI740522:wg16b07.x1 Homo sapiens cDNA, 3 end /clone=IMAGE-  
2365237 /clone\_end=3 /gb=AI740522
- 2 M21154 /FEATURE=mRNA /DEFINITION=HUMAMD Human S-  
adenosylmethionine decarboxylase mRNA, complete cds
- 2 X00737 /FEATURE=cds /DEFINITION=HSPNP Human mRNA for purine  
nucleoside phosphorylase (PNP; EC 2.4.2.1)
- 30 2.1 Cluster Incl. AF034956:Homo sapiens RAD51D mRNA, complete cds  
/cds=(124,993) /gb=AF034956 /gi=2920581
- 2.1 Ras Inhibitor Inf

- 2.1 Cluster Incl. M27749:Human immunoglobulin-related 14.1 protein mRNA, complete cds /cds=(118,759) /gb=M27749
- 2.1 Ras-Like Protein Tc4
- 2.1 X92106 /FEATURE=cds /DEFINITION=HSBLEO H.sapiens mRNA for  
5 bleomycin hydrolase
- 2.1 D88674 /FEATURE= /DEFINITION=D88674 Homo sapiens mRNA for antizyme inhibitor, complete cds
- 2.1 Cluster Incl. H15872:ym22b12.r1 Homo sapiens cDNA, 5 end /clone=IMAGE-48838 /clone\_end=5 /gb=H15872
- 10 2.1 Cluster Incl. L07541:Human replication factor C, 38-kDa subunit mRNA, complete cds /cds=(9,1079) /gb=L07541
- 2.1 V01512 /FEATURE=mRNA#1 /DEFINITION=HSCFOS Human cellular oncogene c-fos (complete sequence)
- 2.1 Cluster Incl. L23959:Homo sapiens E2F-related transcription factor (DP-1) mRNA,  
15 complete cds /cds=(37,1269)
- 2.1 Stimulatory Gdp/Gtp Exchange Protein For C-Ki-Ras P21 And Smg P21
- 2.1 Cluster Incl. L13943:Human glycerol kinase (GK) mRNA exons 1-4, complete cds /cds=(66,1640) /gb=L13943 /gi=348166
- 2.1 Cluster Incl. X78925:H.sapiens HZF2 mRNA for zinc finger protein /cds=(0,2198)  
20 /gb=X78925 /gi=498722 /ug=Hs.2480
- 2.1 X74794 /FEATURE=cds /DEFINITION=HSP1CDC21 H.sapiens P1-Cdc21 mRNA
- 2.1 U78733 /FEATURE=mRNA#1 /DEFINITION=HSSMAD2S8 Homo sapiens mad protein homolog Smad2 gene, exon 11
- 25 2.2 Cluster Incl. L07540:Human replication factor C, 36-kDa subunit mRNA, complete cds /cds=(9,1031) /gb=L07540
- 2.2 Cluster Incl. AF040958:Homo sapiens lysosomal neuraminidase precursor, mRNA, complete cds /cds=(129,1376)
- 2.2 D00596 /FEATURE=cds /DEFINITION=HUMTS1 Homo sapiens gene for  
30 thymidylate synthase, exons 1, 2, 3, 4, 5, 6, 7,
- 2.2 Cluster Incl. AI659108:tu08c09.x1 Homo sapiens cDNA, 3 end /clone=IMAGE-2250448 /clone\_end=3 /gb=AI659108

- 2.2 Cluster Incl. AF042083:Homo sapiens BH3 interacting domain death agonist (BID) mRNA, complete cds /cds=(140,727)
- 2.2 Cluster Incl. W28907:53e12 Homo sapiens cDNA /gb=W28907 /gi=1308855 /ug=Hs.111429 /len=989
- 5 2.3 Cluster Incl. AF073362:Homo sapiens endo/exonuclease Mre11 (MRE11A) mRNA, complete cds /cds=(0,2126)
- 2.3 Escherichia coli /REF=J04423 /DEF=E coli bioD gene dethiobiotin synthetase /LEN=676 (-5 and -3 represent transcript)
- 2.3 Cluster Incl. D59253:Human mRNA for NCBP interacting protein 1, complete cds
- 10 /cds=(36,506) /gb=D59253
- 2.3 M21154 /FEATURE=mRNA /DEFINITION=HUMAMD Human S-adenosylmethionine decarboxylase mRNA, complete cds
- 2.3 Proto-Oncogene C-Myc, Alt. Splice 3, Orf 114
- 2.3 Cluster Incl. W26787:15d8 Homo sapiens cDNA /gb=W26787 /gi=1306078
- 15 /ug=Hs.195188 /len=768
- 2.4 L12002 /FEATURE= /DEFINITION=HUMITGA4A Human integrin alpha 4 subunit mRNA, complete cds
- 2.4 Cluster Incl. M55536:Human glucose transporter pseudogene /cds=UNKNOWN /gb=M55536 /gi=183299 /ug=Hs.121583 2.4 X98743 /FEATURE=cds
- 20 /DEFINITION=HSRNAHELC H.sapiens mRNA for RNA helicase (Myc-regulated dead box protein)
- 2.4 S75881 /FEATURE= /DEFINITION=S75881 A-myb=DNA-binding transactivator {3 region} [human, CCRF-CEM T leukemia line, mRNA Partial, 831 nt]
- 25 2.4 Cluster Incl. AF050110:Homo sapiens TGFb inducible early protein and early growth response protein alpha genes, complete cds /cds=(123,1565) /gb=AF050110 /gi=3523144 /ug=Hs.82173 /len=2899
- 2.5 Cluster Incl. M86667:H.sapiens NAP (nucleosome assembly protein) mRNA, complete cds /cds=(75,1250) /gb=M86667
- 30 /gi=189066 /ug=Hs.179662 /len=1560

- 2.5 U17743 /FEATURE= /DEFINITION=HSU17743 Human JNK activating kinase (JNKK1) mRNA, complete cds
- 2.5 Cluster Incl. U90549:Human non-histone chromosomal protein (NHC) mRNA, complete cds /cds=(691,963) /gb=U90549
- 5 /gi=2062699 /ug=Hs.63272 /len=1981
- 2.5 Cluster Incl. U31382:Human G protein gamma-4 subunit mRNA, complete cds /cds=(98,325) /gb=U31382 /gi=995916
- /ug=Hs.32976 /len=670
- 2.5 Cluster Incl. S81916:phosphoglycerate kinase {alternatively spliced} [human, phosphoglycerate kinase deficient patient
- 10 with episodes of muscl, mRNA Partial Mutant, 307 nt] /cds=(0,143) /gb=S81916 /gi=1470308 /ug=Hs.169313 /len=307
- 2.5 Cluster Incl. M64595:Human small G protein (Gx) mRNA, 3 end /cds=(0,542) /gb=M64595 /gi=183708 /ug=Hs.173466
- 15 /len=757
- 2.5 Serine Hydroxymethyltransferase, Cytosolic, Alt. Splice 3
- 2.5 U88629 /FEATURE=cds /DEFINITION=HSU88629 Human RNA polymerase II elongation factor ELL2, complete cds
- 2.5 Cluster Incl. U72518:Human destrin-2 pseudogene mRNA, complete cds
- 20 /cds=(268,798) /gb=U72518 /gi=1673523 /ug=Hs.199299 /len=1057
- 2.5 Cluster Incl. L14595:Human alanine/serine/cysteine/threonine transporter (ASCT1) mRNA, complete cds
- 2.5 Cluster Incl. AB014584:Homo sapiens mRNA for KIAA0684 protein, partial cds
- 25 /cds=(0,2711) /gb=AB014584 /gi=3327181 /ug=Hs.24594 /len=4124
- 2.5 Cluster Incl. AI924594:wn57a11.x1 Homo sapiens cDNA, 3 end /clone=IMAGE-2449532 /clone\_end=3 /gb=AI924594
- /gi=5660558 /ug=Hs.122540 /len=685
- 30 2.5 U68111 /FEATURE=mRNA /DEFINITION=HSPPP1R2E6 Human protein phosphatase inhibitor 2 (PPP1R2) gene, exon 6

- 2.5 Cluster Incl. AL009179:dJ97D16.4 (Histone H2B) /cds=(25,405) /gb=AL009179  
/gi=3217024 /ug=Hs.137594 /len=488
- 2.6 Cluster Incl. AF091077:Homo sapiens clone 558 unknown mRNA, complete  
sequence /cds=(1,300) /gb=AF091077  
5 /gi=3859991 /ug=Hs.40368 /len=947
- 2.7 Cluster Incl. M28211:Homo sapiens GTP-binding protein (RAB4) mRNA,  
complete cds /cds=(70,711) /gb=M28211  
/gi=550067 /ug=Hs.234038 /len=735
- 2.6 X69549 /FEATURE=cds /DEFINITION=HSRHO2 H.sapiens mRNA for rho  
10 GDP-dissociation Inhibitor 2
- 2.6 Cluster Incl. Z85986:Human DNA sequence from clone 108K11 on chromosome  
6p21 Contains SRP20 (SR protein family  
member), Ndr protein kinase gene similar to yeast suppressor protein SRP40, EST  
and GSS /cds=(0,932) /gb=Z85986  
15 gi=4034056 /ug=Hs.152400 /len=933
- 2.6 Zinc Finger Protein, Kruppel-Like
- 2.7 D10656 /FEATURE= /DEFINITION=HUMCRK Human mRNA for CRK-II,  
complete cds
- 2.7 M28211 /FEATURE= /DEFINITION=HUMRAB4A Homo sapiens GTP-binding  
20 protein (RAB4) mRNA, complete cds
- 2.7 Cluster Incl. AB019435:Homo sapiens mRNA for putative phospholipase,  
complete cds /cds=(72,3074) /gb=AB019435  
/gi=4760646 /ug=Hs.125670 /len=3088
- 2.8 U39318 /FEATURE= /DEFINITION=HSU39318 Human E2 ubiquitin conjugating  
25 enzyme UbCH5C (UBCH5C) mRNA,  
complete cds
- 2.9 Cluster Incl. X78711:H.sapiens mRNA for glycerol kinase testis specific 1  
/cds=(26,1687) /gb=X78711 /gi=515028  
/ug=Hs.1466 /len=1838
- 30 2.8 Cluster Incl. W27594:34h4 Homo sapiens cDNA /gb=W27594 /gi=1307542  
/ug=Hs.8258 /len=702

- 2.8 X05360 /FEATURE=cds /DEFINITION=HSCDC2 Human CDC2 gene involved in cell cycle control
- 2.8 V00568 /FEATURE=cds /DEFINITION=HSMYC1 Human mRNA encoding the c-myc oncogene
- 5 2.8 Cluster Incl. L24804:Human (p23) mRNA, complete cds /cds=(232,714) /gb=L24804 /gi=438651 /ug=Hs.75839 /len=782
- 2.10 Cluster Incl. Y09443:H.sapiens mRNA for alkyl-dihydroxyacetonephosphate synthase precursor /cds=(15,1991) /gb=Y09443 /gi=1922284 /ug=Hs.22580 /len=2074
- 10 2.8 Cluster Incl. Z82200:Human DNA sequence from clone 333E23 on chromosome Xq21.1 Contains putative purinergic receptor P2Y10 /cds=(0,1019) /gb=Z82200 /gi=2370075 /ug=Hs.166137 /len=1020
- 2.9 L05624 /FEATURE= /DEFINITION=HUMMKK Homo sapiens MAP kinase kinase mRNA, complete cds
- 15 2.9 Cluster Incl. D88357:Homo sapiens mRNA for CDC2 delta T, complete cds /cds=(27,749) /gb=D88357 /gi=3126638 /ug=Hs.184572 /len=780
- 20 Up regulated genes due to LQGV treatment  
Fold Change/Descriptions
- 4.9 Cluster Incl. AF043324:Homo sapiens N-myristoyltransferase 1 mRNA, complete cds (cds=(10,1500)/gb=AF043324 /gi=3005062 /ug=Hs.111039 /len=4378)
- 25 3.3 Cluster Incl. L08096:Human CD27 ligand mRNA, complete cds /cds=(150,731) (gb=L08096 /gi=307127 /ug=Hs.99899 /len=926)
- 30 2 Cluster Incl. AF043325:Homo sapiens N-myristoyltransferase 2 mRNA, complete cds /cds=(46,1542) /gb=AF043325 /gi=3005064 /ug=Hs.122647 /len=2838



- 2.1 Cluster Incl. AL031681:dJ862K6.2.2 (splicing factor, arginine/serine-rich 6 (SRP55-2)(isoform 2)) /cds=(106,513)
- 2.1 Cluster Incl. X87838:H.sapiens mRNA for beta-catenin /cds=(214,2559) /gb=X87838 /gi=1154853 /ug=Hs.171271
- 5 2.2 Cluster Incl. AW024285:wt69d06.x1 Homo sapiens cDNA, 3 end /clone=IMAGE-2512715 /clone\_end=3 /gb=AW024285
- 2.2 Cluster Incl. D38524:Human mRNA for 5-nucleotidase /cds=(83,1768) /gb=D38524 /gi=633070 /ug=Hs.138593
- 2.2 Cluster Incl. L38935:Homo sapiens GT212 mRNA /cds=UNKNOWN /gb=L38935
- 10 /gi=1008845 /ug=Hs.83086 /len=1165
- 2.5 Cluster Incl. L12711:Homo sapiens transketolase (tk) mRNA, complete cds /cds=(98,1969) /gb=L12711 /gi=388890
- 2.6 Cluster Incl. AF026029:Homo sapiens poly(A) binding protein II (PABP2) gene, complete cds /cds=(1282,2202)
- 15 2.8 Cluster Incl. X70683:H.sapiens mRNA for SOX-4 protein /cds=(350,1774) /gb=X70683 /gi=36552 /ug=Hs.83484

#### Further examples of use

20 Examples of different receptor-intracellular signaling pathways involved in different disease pathogenesis where signaling molecules according to the invention find their use are:

LPS stimulation of antigen presenting cells (like DC, macrophages, monocytes) through different Toll-like receptors activates different signaling pathways including, MAPK pathways, ERK, JNK and p38 pathways. These pathways directly or indirectly phosphorylate and activate various transcription factors, including Elk-1, c-Jun, c-Fos, 25 ATF-1, ATF-2, SRF, and CREB. In addition, LPS activates the IKK pathway of MyD88, IRAK, and TRAF6. TAK1-TAB2 and MEKK1-ECSIT complexes phosphorylate IKKb, which in turn phosphorylates IkbBs. Subsequent degradation of IkbBs permits nuclear translocation of NFkB/Rel complexes, such as p50/p65. Moreover, the P13K-Akt pathway 30 phosphorylates and activates p65 via an unknown kinase. Some of these pathways could also be regulated by other receptor signaling molecules such as hormones/growth factor receptor tyrosine kinases (PKC/Ras/IRS pathway) and cytokine receptors (JAK/STAT

pathway). In the genomic experiment with the T-cell line several of these genes appeared to be downregulated or upregulated by the peptide used (LQGV). It is now clear that other peptides in T cells and the same and other peptides in other cell types similarly down-regulate or up-regulate several of these transcription factors and signaling molecules. In  
5 DC and fertilized eggs experiments NMPF had the ability to modulate growth factor (GM-CSF, VEGF) and LPS signaling. Some diseases associated with dysregulation of NF-kB and related transcription factors are: Atherosclerosis, asthma, arthritis, anthrax, cachexia, cancer, diabetes, euthyroid sick syndrome, AIDS, inflammatory bowel disease, stroke, (sepsis) septic shock, inflammation, neuropathological diseases, autoimmunity,  
10 thrombosis, cardiovascular disease, psychological disease, post surgical depression, wound healing, burn-wounds healing and neurodegenerative disorders.

PKC plays an essential role in T cell activation via stimulation of for example AP-1 and NF-kB that selectively translocate to the T cell synapse via Vav/Rac pathway. PKC is involved in a variety of immunological and non-immunological diseases as is clear from  
15 standard text books of internal medicine (examples are metabolic diseases, cancer, angiogenesis, immune mediated disorders, diabetes etc.)

LPS and ceramide induce differential multimeric receptor complexes, including CD14, CD11b, Fc-gRIII, CD36, TAPA, DAF and TLR4. This signal transduction pathway explains the altered function of monocytes in hypercholesterolemia and lipid disorders.

20 Oxidized low-density lipoproteins contribute to stages of the atherogenic process and certain concentrations of oxidized low-density lipoproteins induce apoptosis in macrophages through signal transduction pathways. These pathways are involved in various vascular diseases such as atherosclerosis, thrombosis etc.

Bacterial DNA is recognized by cells of the innate immune system. This  
25 recognition requires endosomal maturation and leads to activation of NF-kB and the MAPK pathway. Recently it has been shown that signaling requires the Toll like receptor 9 (TLR9) and the signaling adaptor protein MyD88. Recognition of dsRNA during viral infection seems to be dependent on intracellular recognition by the dsRNA dependent protein kinase PKR. TLRs play an essential role in the immune system and they are  
30 important in bridging and balancing innate immunity and adaptive immunity. Modulation of these receptors or their down-stream signaling pathways are important for the treatment of various immunological conditions such as infections, cancer, immune-mediated

diseases, autoimmunity, certain metabolic diseases with immunological component, vascular diseases, inflammatory diseases etc.

Effect of growth factor PDGF-AA on NF-kB and proinflammatory cytokine expression in rheumatoid synoviocytes; PDGF-AA augmented NF-kB activity and mRNA expression of IL-1b, IL-8 and MIP-1a. Therefore, PDGF-AA may play an important role in progression of inflammation as well as proliferation of synoviocytes in RA.

Dendritic cell (DC) activation is a critical event for the induction of immune responses. DC activation induced by LPS can be separated into two distinct processes: first, maturation, leading to upregulation of MHC and costimulatory molecules, and second, rescue from immediate apoptosis after withdrawal of growth factors (survival). LPS induces NF-kB transcription factor. Inhibition of NF-kB activation blocked maturation of DCs in terms of upregulation of MHC and costimulatory molecules. In addition, LPS activates the extracellular signal-regulated kinases (ERK), and specific inhibition of MEK1, the kinase which activates ERK, abrogate the ability of LPS to prevent apoptosis but do not inhibit DC maturation or NF-kB nuclear translocation. This shows that ERK and NF-kB regulate different aspects of LPS induced DC activation. Our DC data and NF-kB data also show the various effects of NMPF peptide on DC maturation and proliferation in the presence or absence of LPS. NMPF peptides modulate these pathways and are novel tools for the regulation of DC function and immunoregulation. This opens new ways for the treatment of immune diseases, particularly those in which the immune system is in disbalance (DC1-DC2, Th1-Th2, regulatory cell etc.)

DC mediate NK cell activation which can result in tumor growth inhibition. DC cells and other antigen presenting cells (like macrophages, B-cells) play an essential role in the immune system and they are also important in bridging and balancing innate immunity and adaptive immunity. Modulation of these cells or their down-stream signaling pathways are important for the treatment of various immunological conditions such as infections, cancer, immune-mediated diseases, autoimmunity, certain metabolic diseases with immunological component, vascular diseases, inflammatory diseases etc. There is also evidence in the literature that mast cells play important roles in exerting the innate immunity by releasing inflammatory cytokines and recruitment of neutrophils after recognition of infectious agents through TLRs on mast cells.

In murine macrophages infected with *Mycobacterium tuberculosis* through JAK pathway activate STAT1 and activation of STAT1 may be the main transcription factor involved in IFN-g-induced MHC class II inhibition.

Recognition of mannose-binding lectin (MBL) through TLRs influences multiple immune mechanisms in response to infection and involved in innate immunity. Balance between innate and adoptive immunity is crucial for balanced immune system and dysregulation in immune system lead to different spectrum of diseases such as, inflammatory diseases, autoimmunity, infectious diseases, pregnancy associated diseases (like miscarriage and pre-eclampsia), diabetes, atherosclerosis and other metabolic diseases.

Nuclear factor-kappaB (NFkappaB) is critical for the transcription of multiple genes involved in myocardial ischemia-reperfusion injury. Clinical and experimental studies have shown that myocardial ischemia-reperfusion injury results in activation of the TLRs and the complement system through both the classical and the alternative pathway in myocardial infarction, atherosclerosis, intestinal ischaemia, hemorrhagic shock pulmonary injury, and cerebral infarction etc.

Peroxisome proliferator-activated receptors (PPARs) are ligand-activated transcription factors which function as regulators of lipid and lipoprotein metabolism, glucose homeostasis, influence cellular proliferation, differentiation and apoptosis and modulation of inflammatory responses. PPAR alpha is highly expressed in liver, muscle, kidney and heart, where it stimulates the beta-oxidative degradation of fatty acids. PPAR gamma is predominantly expressed in intestine and adipose tissue, where it triggers adipocyte differentiation and promotes lipid storage. Recently, the expression of PPAR alpha and PPAR gamma was also reported in cells of the vascular wall, such as monocyte/macrophages, endothelial and smooth muscle cells. The hypolipidemic fibrates and the antidiabetic glitazones are synthetic ligands for PPAR alpha and PPAR gamma, respectively. Furthermore, fatty acid-derivatives and eicosanoids are natural PPAR ligands: PPAR alpha is activated by leukotriene B4, whereas prostaglandin J2 is a PPAR gamma ligand, as well as of some components of oxidized LDL, such as 9- and 13-HODE. These observations suggested a potential role for PPARs not only in metabolic but also in inflammation control and, by consequence, in related diseases such as atherosclerosis. More recently, PPAR activators were shown to inhibit the activation of inflammatory

response genes (such as IL-2, IL-6, IL-8, TNF alpha and metalloproteases) by negatively interfering with the NF-kappa B, STAT and AP-1 signaling pathways in cells of the vascular wall. Furthermore, PPARs may also control lipid metabolism in the cells of the atherosclerotic plaque. PPARs are also involved in a variety of immunological and non-immunological diseases as is clear from standard text books of internal medicine (examples are metabolic diseases, cancer, angiogenesis, immune mediated disorders, diabetes etc.)

As mentioned above the nuclear receptor PPAR $\gamma$  is important in adipogenesis, lipid storage and involved in atherosclerosis. While expressed in adipose tissue this receptor is also expressed in macrophages and in the colon. In addition, PPAR $\gamma$  is implicated in a number of processes such as cancer and inflammation. Moreover, microbes, via its cognate receptors, typified by the TLRs, possess the capacity to regulate PPAR $\gamma$  dependent metabolic functions and as such illustrates the intricate interplay between the microbial flora and metabolic control in the alimentary tract.

Cyclo-oxygenase 2 (COX2), an inducible isoform of prostaglandin H synthase, which mediates prostaglandin synthesis during inflammation, and which is selectively overexpressed in colon tumors, is thought to play an important role in colon carcinogenesis. Induction of COX2 by inflammatory cytokines or hypoxia-induced oxidative stress can be mediated by nuclear factor kappa B (NF-kappaB). So, inhibition of NF-kB modulate COX pathway and this inhibition of NF-kB can be therapeutically useful in diseases in which COXs are involved, such as inflammation, pain, cancer (especially colorectal cancer), inflammatory bowel disease and others.

Neuronal subsets in normal brains constitutively express functionally competent C5a receptors. The functional role of C5a receptors revealed that C5a triggered rapid activation of protein kinase C and activation and nuclear translocation of the NF-kappa B transcription factor. In addition, C5a was found to be mitogenic for undifferentiated human neuroblastoma cells, a novel action for the C5aR. In contrast, C5a protects terminally differentiated human neuroblastoma cells from toxicity mediated by the amyloid A beta peptide. This shows that normal hippocampal neurons as well as undifferentiated and differentiated human neuroblastoma cells express functional C5a receptors. These results show the role of neuronal C5aR receptors in normal neuronal development, neuronal homeostasis, and neuroinflammatory conditions such as Alzheimer's disease.

Activation of the complement system plays also an important role in the pathogenesis of atherosclerosis. The proinflammatory cytokine interleukin (IL)-6 is potentially involved in the progression of the disease. Here the complement system induces IL-6 release from human vascular smooth-muscle cells (VSMC) by a Gi-dependent pathway involving the generation of oxidative stress and the activation of the redox sensitive transcription factors NF-kB and AP-1. Modulation of complement system is important for broad ranges of disorders such as blood disorders, infections, some metabolic diseases (diabetes), vascular diseases, transplant rejection and related disorders, autoimmune diseases, and other immunological diseases.

Different transcription factors like NF-kB and intracellular signaling molecules such as different kinases are also involved in multiple drug resistance. So, it is reasonable to believe that NMPF peptides will be effective against multiple drug resistance. Moreover, our genomic data shows that a number of genes and signaling molecules involved in tumorogenesis and metastasis are modulated. In addition since oligopeptides have also effect on angiogenesis, thus these peptides will also be used for the treatment of cancer and related diseases whereby angiogenesis requires modulation.

Proliferative diabetic retinopathy (PDR) is one of the major causes of acquired blindness. The hallmark of PDR is neovascularization (NV), abnormal angiogenesis that may ultimately cause severe vitreous cavity bleeding and/or retinal detachment. Since NMPF peptides have angiogenesis stimulatory as well as inhibitory effects and have the ability to modulate intracellular signaling involved in growth factors (like insulin), pharmacologic therapy with certain NMPF peptides can improve metabolic control (like glucose) or blunt the biochemical consequences of hyperglycaemia (through mechanisms such as in which aldose reductase, protein kinase C (PKC), PPARs are involved). For this metabolic control or diabetes (type 2) NMPF (LQGV, VLPALP, VLPALPQ, GVLPALPQ, AQQ, LAG, LQA, AQGV, VAPALP, VAPALPQ, VLPALPA, LPGC, MTR, MTRV, LQG, CRGVNPVVS are recommended. The angiogenesis in PDR could be also treated with above mentioned oligopeptides.